

2023 금연 연구회 제7차 심포지엄

# 쉽게 배우는 유세포 분석기법과 담배추출물을 활용한 실험예시

충남대학교병원 호흡기내과

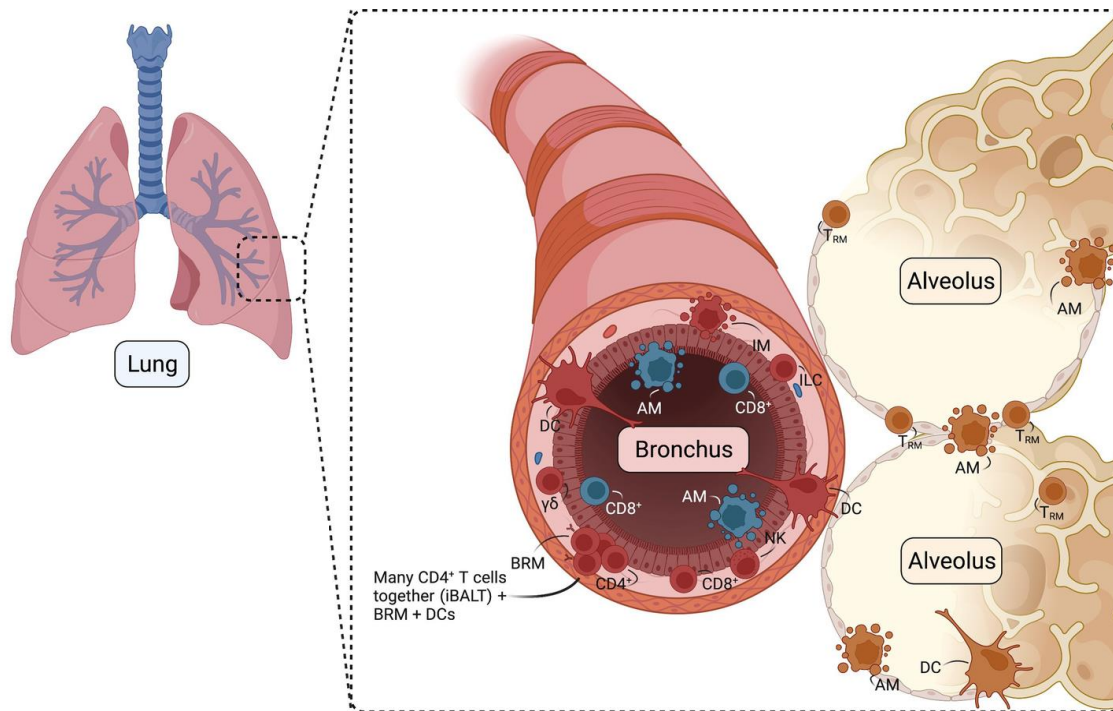
김덕기

# 유세포 분석(flow cytometry)

: An essential tool for immunological studies

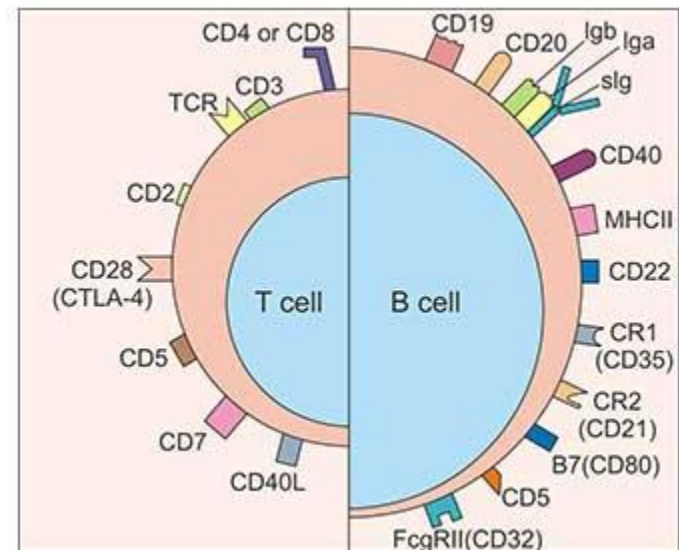
## Flow cytometry

: The measurement (-metry) of cell (cyto-) characteristics and properties in motion (flow)



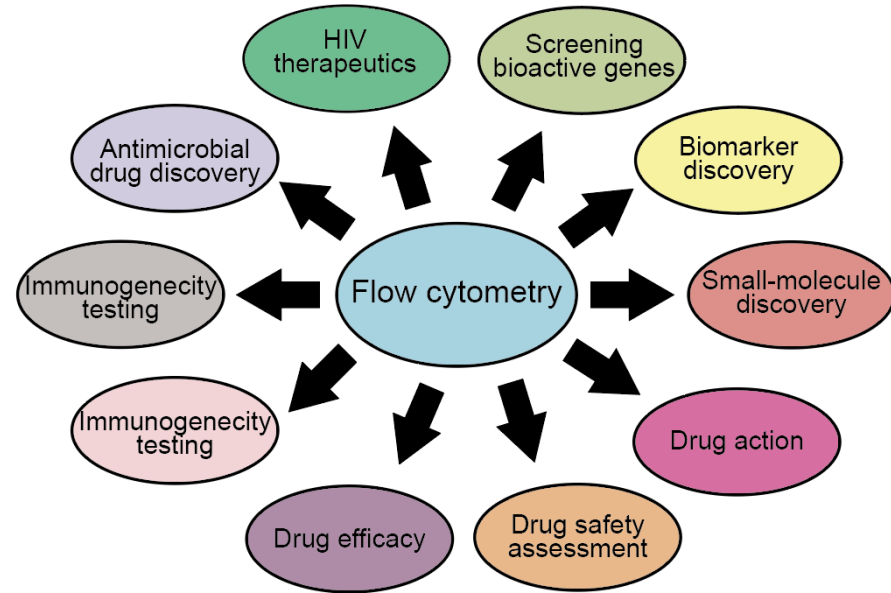
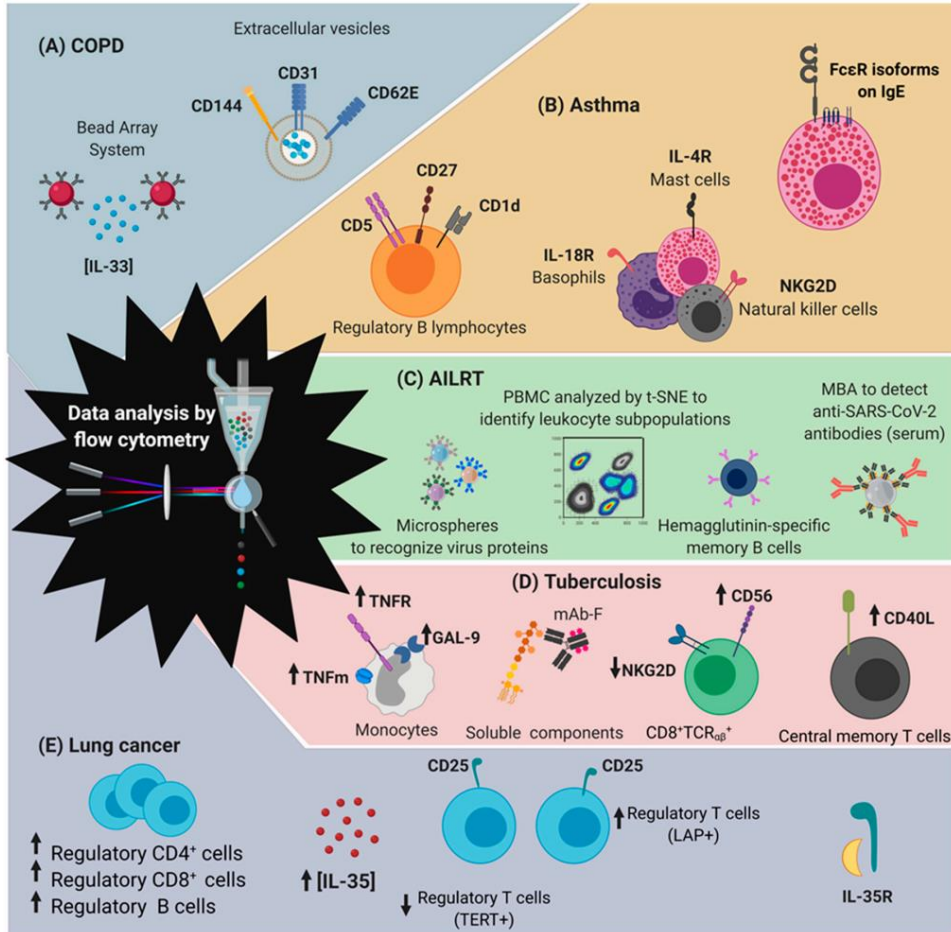
Zazara et al., 2022, *Semin. Immunopathol.*

Each type of immune cell differentially expresses marker proteins



... allows for the identification and characterization of immune cell subsets within a heterogenous population...!

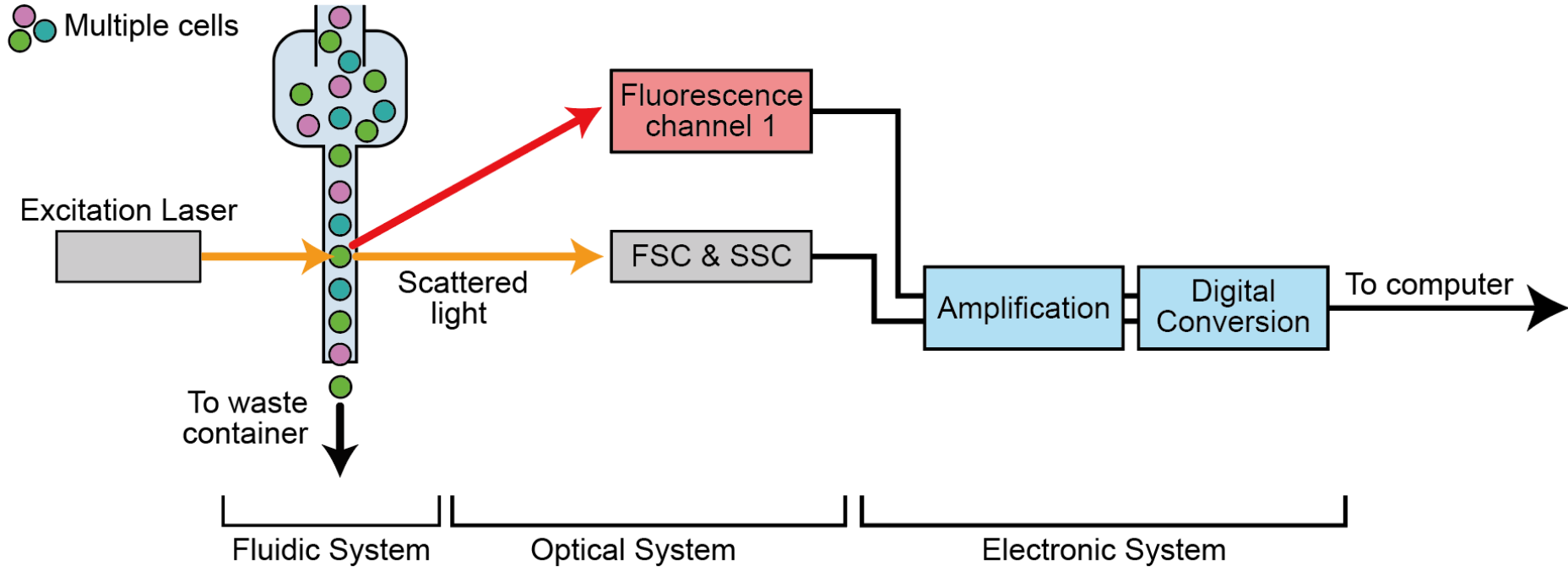
# 유세포 분석의 활용 범위



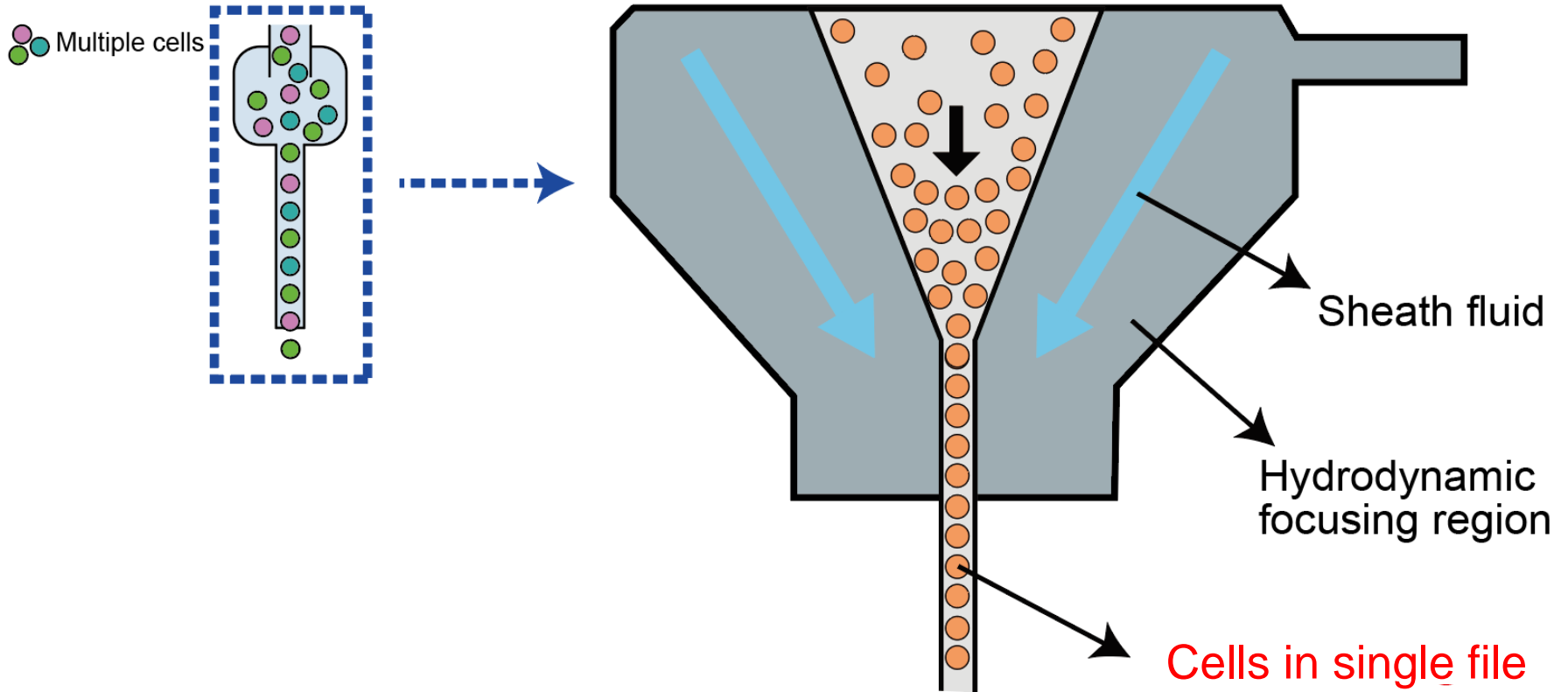
*Julio et al., 2020, Int. J. Mol. Sci.*

# 유세포 분석의 작동 원리

→ 유세포 분석은 단일 또는 다중 레이저를 사용하여 (형광이 표시된) 단일 세포의 다중 파라미터 분석을 하는 기법입니다.

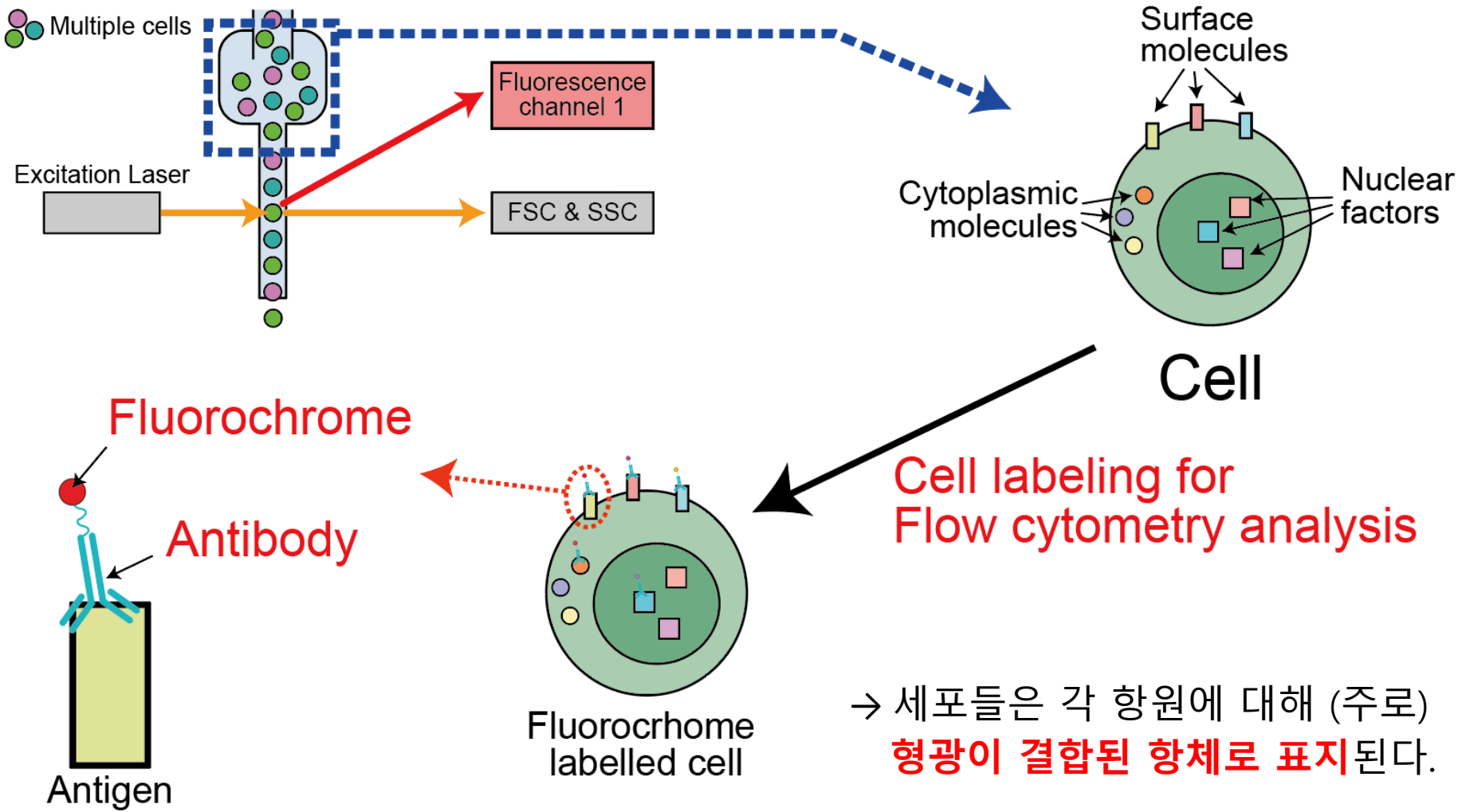


# 유세포 분석의 작동 원리 - Fluidic system

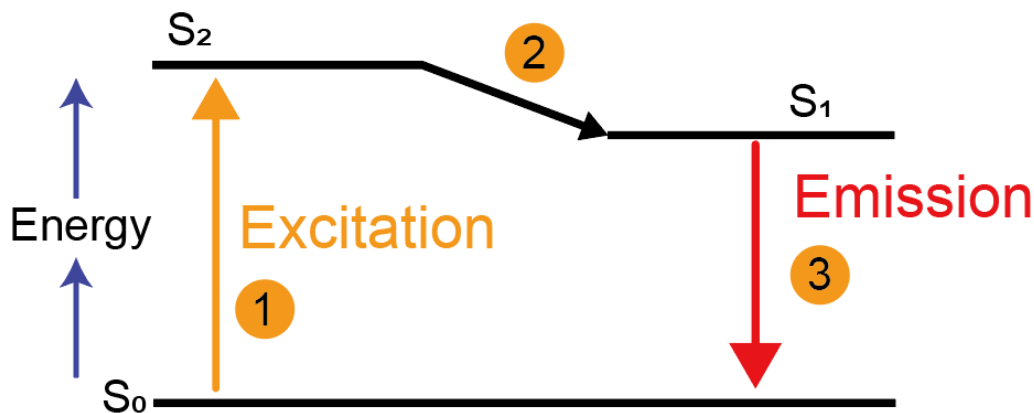
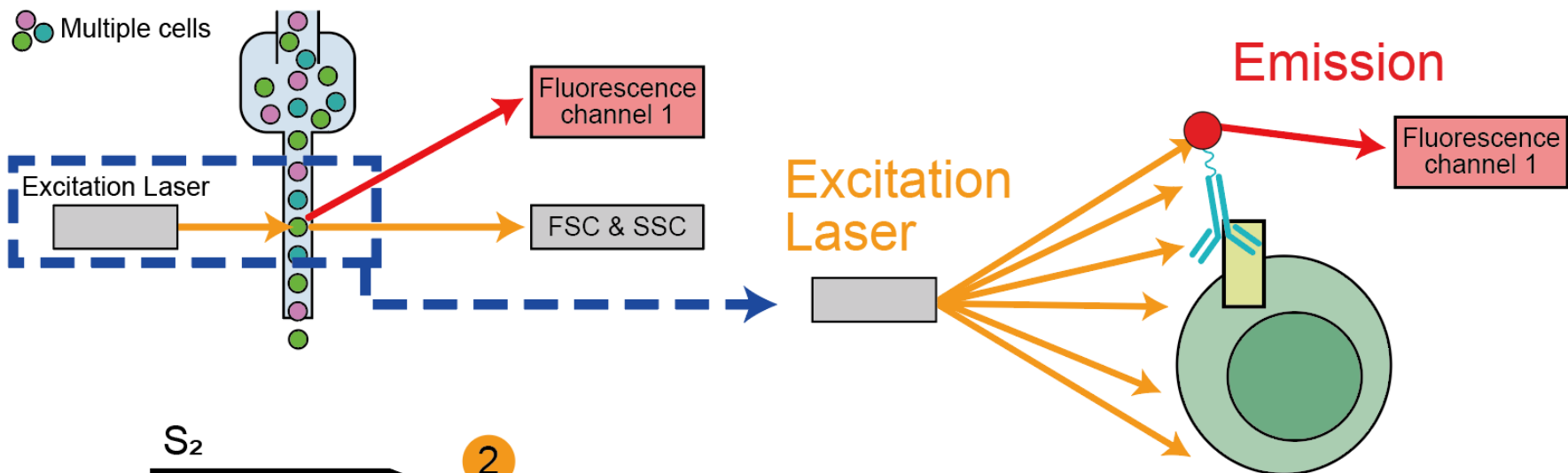


→ Fluidic system은 세포 하나하나의 데이터를 얻게 만들어준다.

# 유세포 분석의 작동 원리 - Cell Labeling

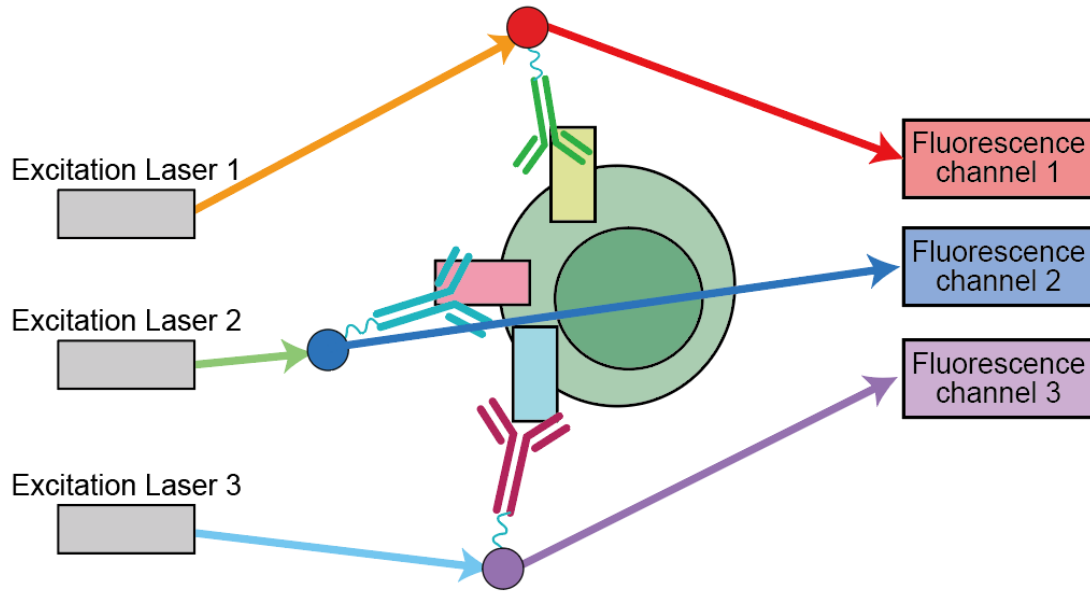


# 유세포 분석의 작동 원리 - Excitation & Emission

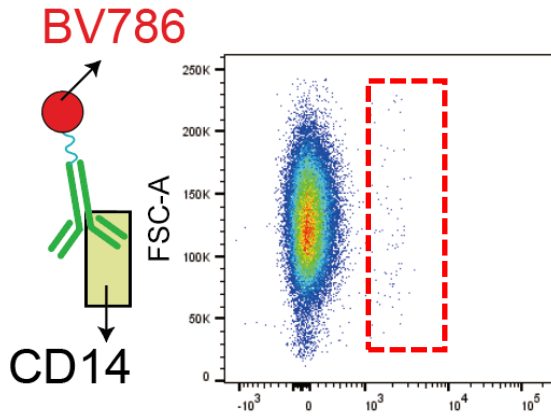


→ 결국, 세포에 표지된 **형광을 읽어서 기록**하는 것이다.

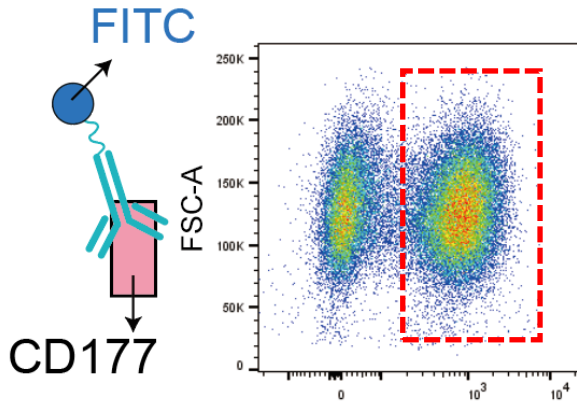
# 유세포 분석의 작동 원리 - Multiple Fluorochrome



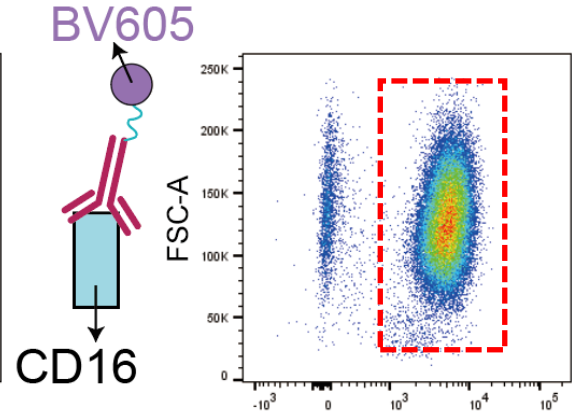
세포가 발현하는 여러가지 항원에 대해 **각기 다른 형광이 결합된 항체를 표지**하여, **각각의 형광을 서로 다른 채널에서** 값을 읽어내 나타낸다.



BV786

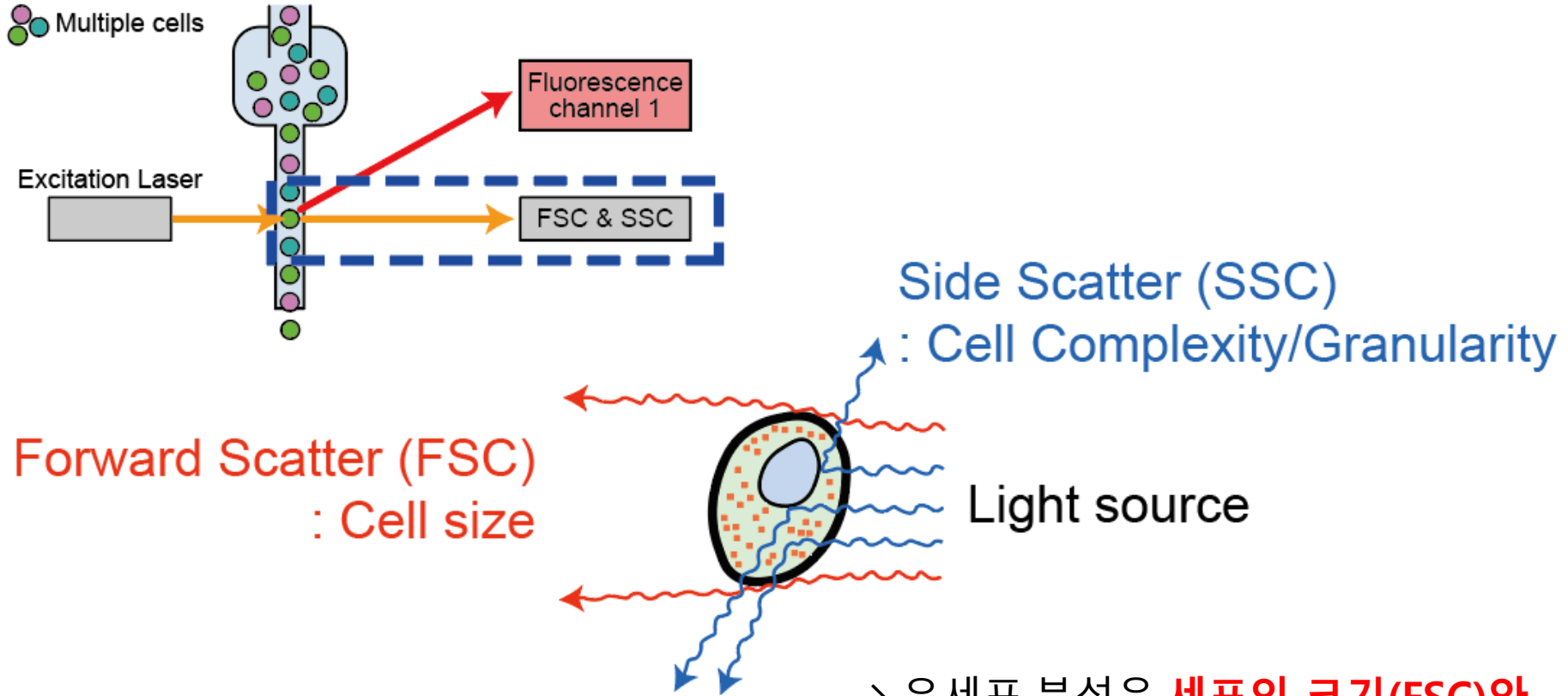


FITC



BV605

# 유세포 분석의 작동 원리 - Morphologic Gating

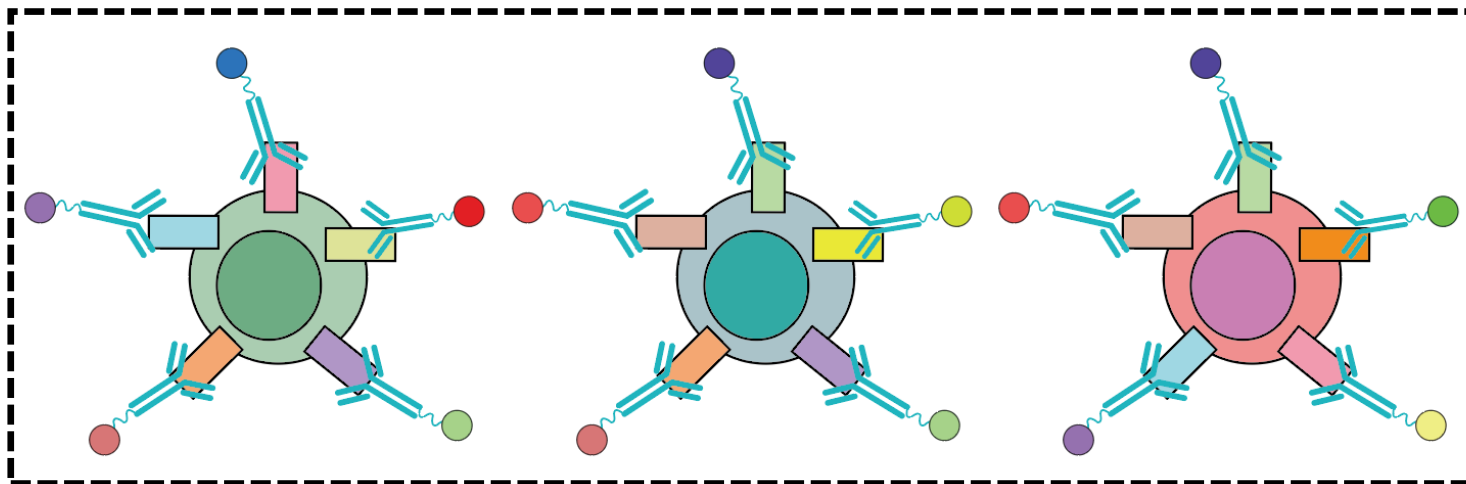


Forward Scatter (FSC)  
: Cell size

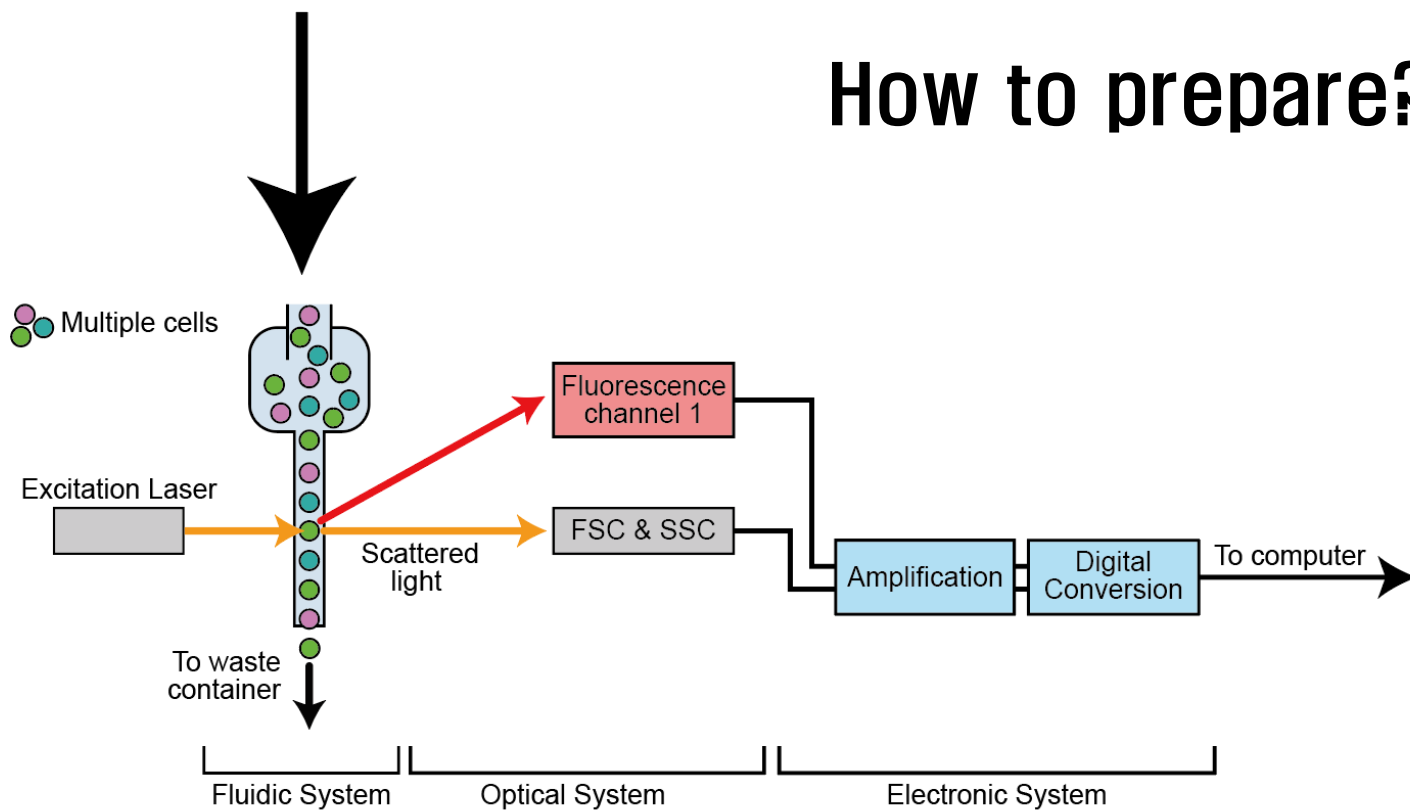
Side Scatter (SSC)  
: Cell Complexity/Granularity

→ 유세포 분석은 **세포의 크기(FSC)**와 **복잡성(SSC)**을 활용하여 형태학적 구분이 가능하다.

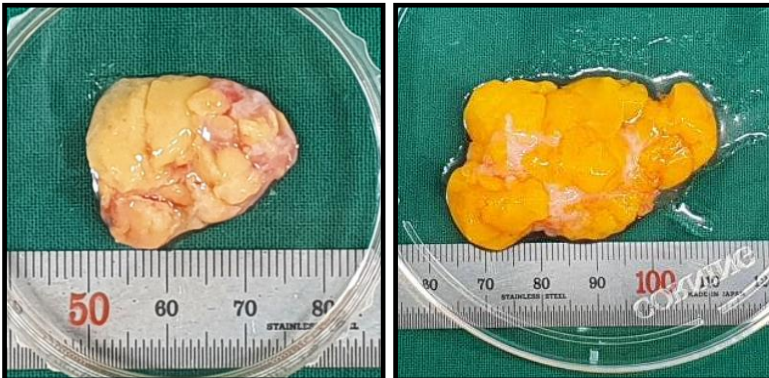
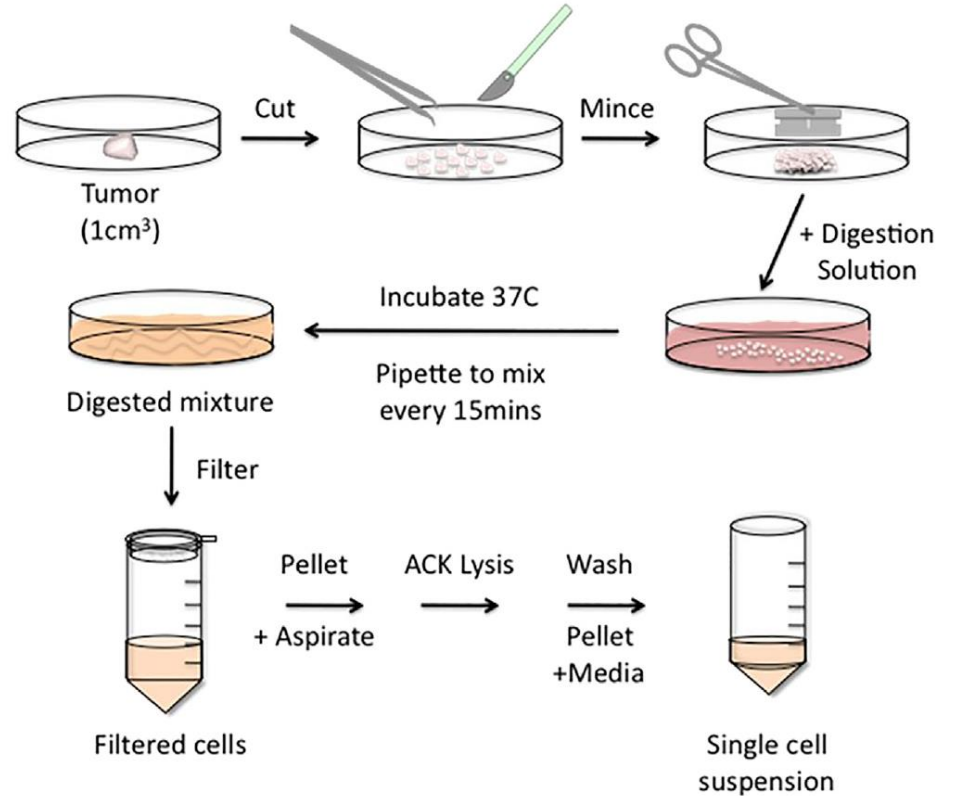
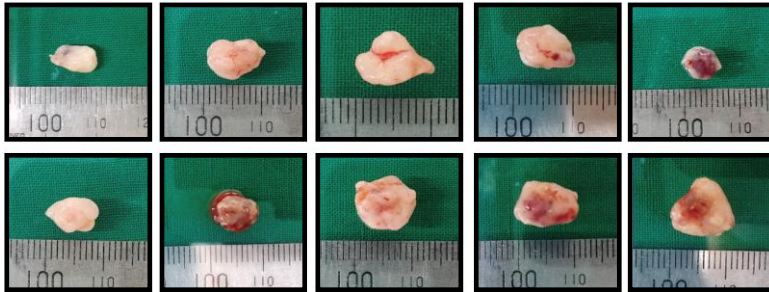
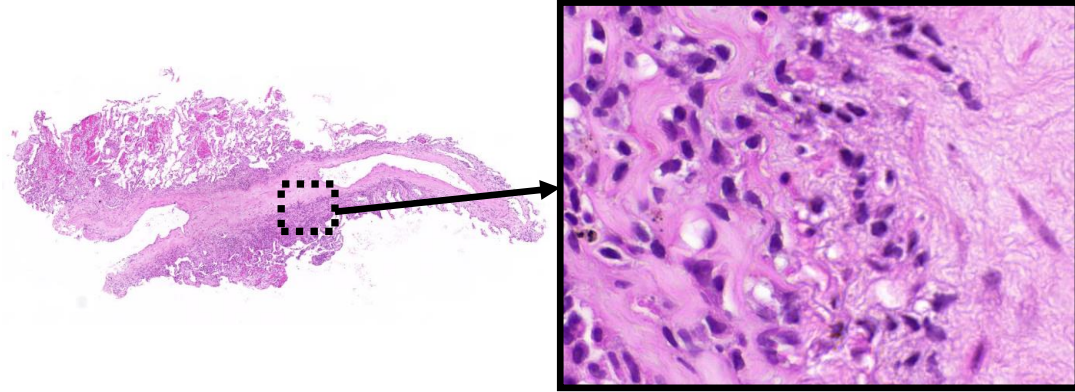
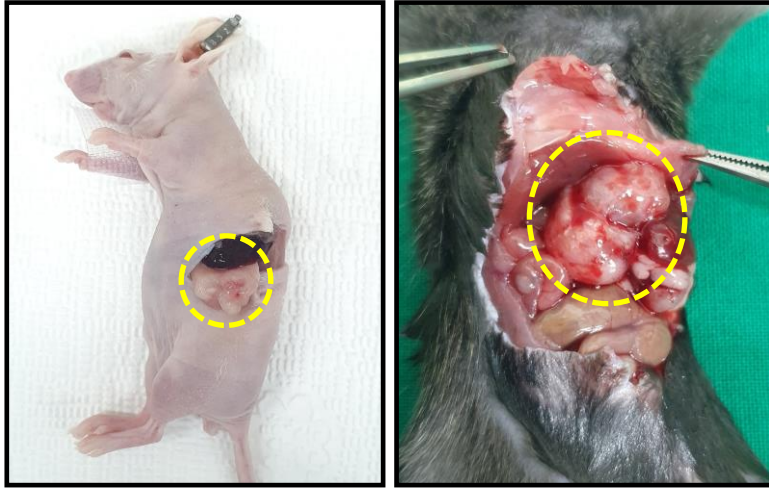
# 유세포 분석을 위한 세포 준비 과정



How to prepare?



# 유세포 분석을 위한 세포 준비 - 조직(tissue)



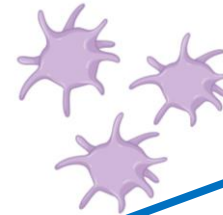
# 유세포 분석을 위한 세포 준비 - 혈액(blood)



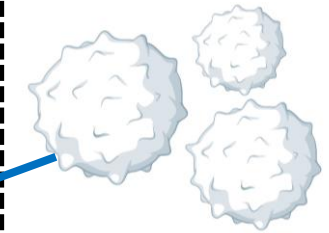
Red Blood Cells



Platelets

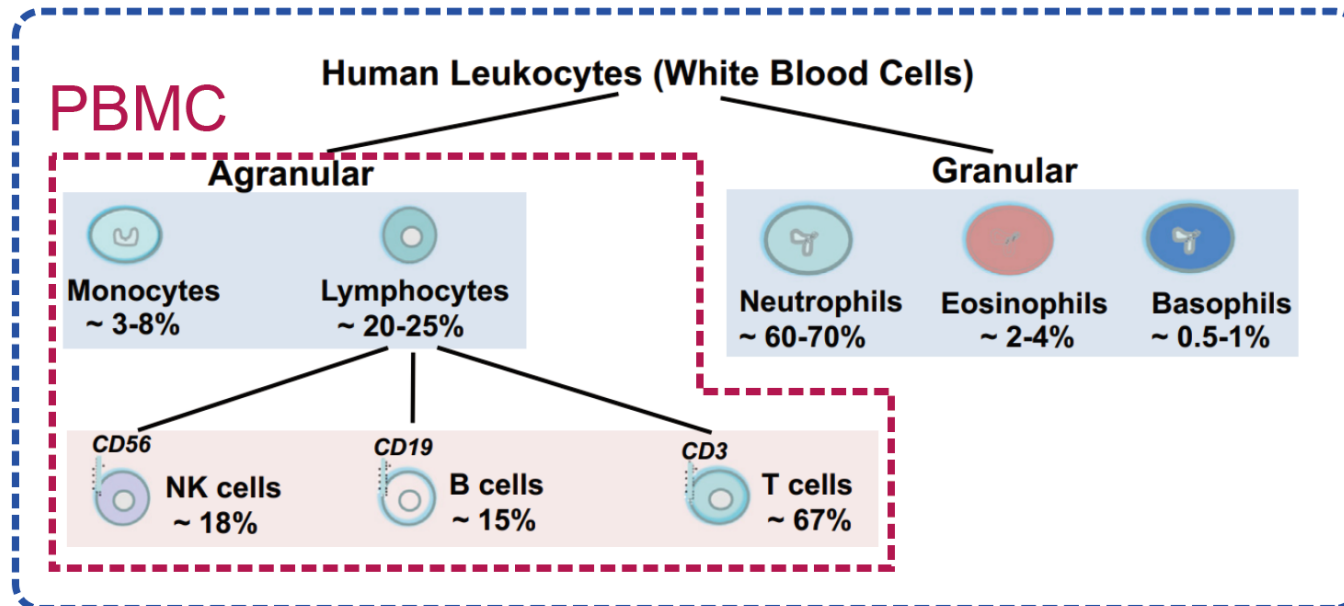


White Blood Cells  
(Leukocytes)



*Cited from freepik, brgfx*

## Whole blood analysis

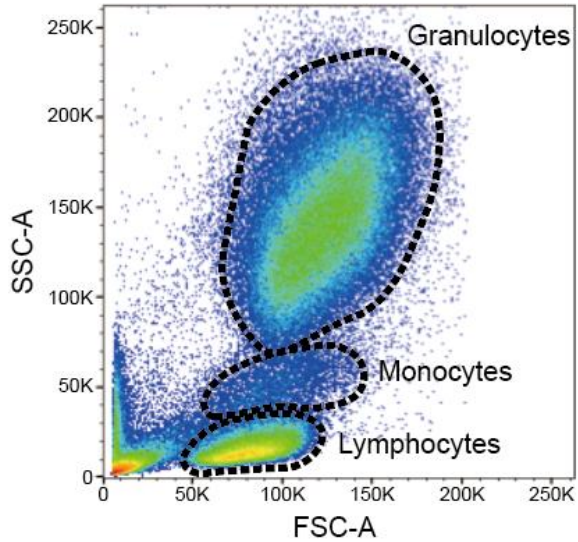


*Cited from Personalized Epigenetics, 2015*

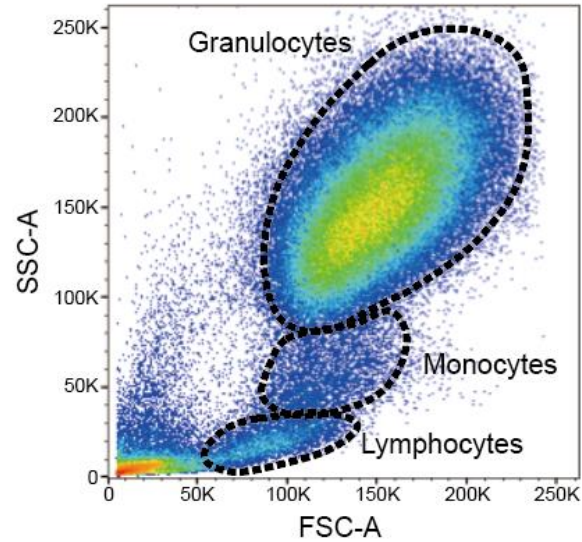
# Whole blood vs PBMC

## Whole blood analysis

Healthy control

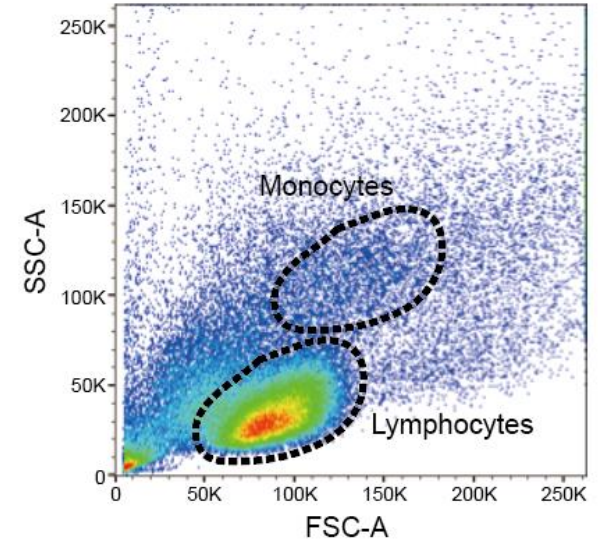


Sepsis patient

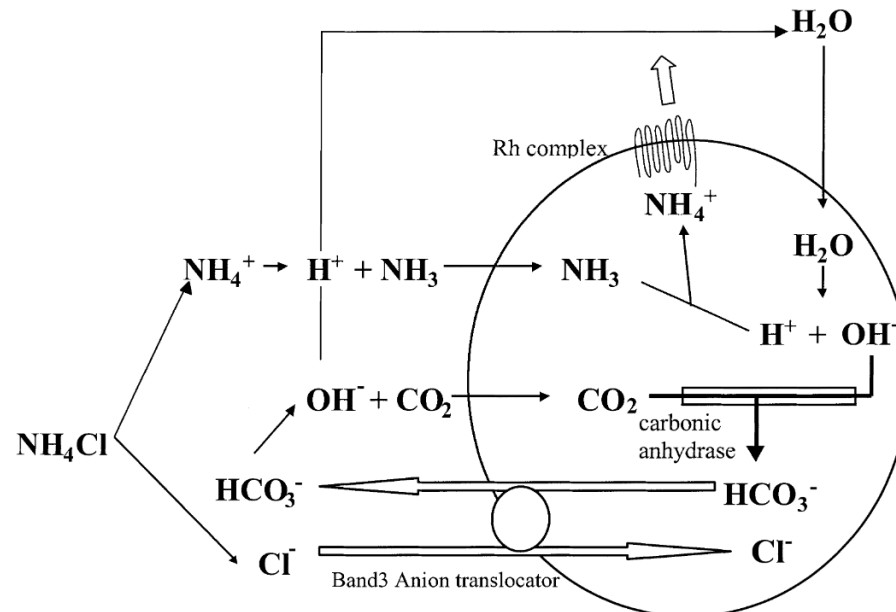
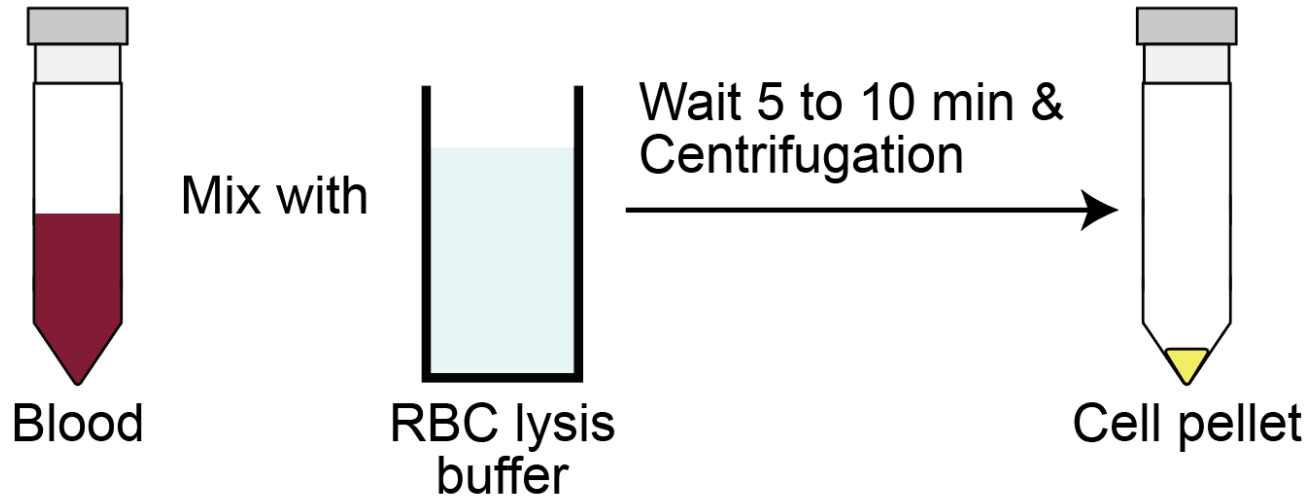


## PBMC isolation

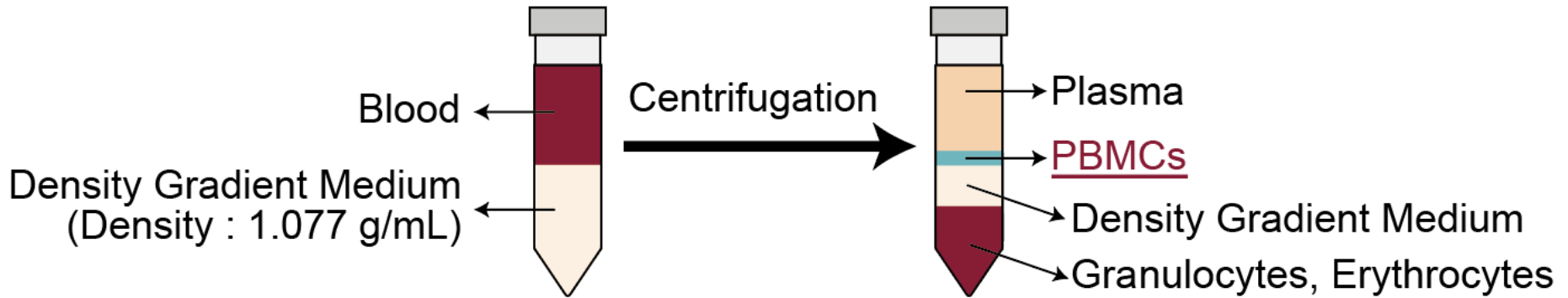
Healthy control



# Whole blood preparation



# PBMC preparation



**Table 1. The specific densities and effective radii of common blood components.**

<b>Blood cell type</b>	<b>Specific density(g/mL)</b>	<b>Effective radius (<math>\mu\text{m}</math>)</b>
Monocyte	1.067–1.077[22]	6–7.5[23]
Lymphocyte	1.073–1.077[22]	3–6[23]
Basophil	1.072–1.078[24]	4.5–5[23]
Neutrophil	1.08–1.09[25]	6–7.5[23]
Eosinophil	1.09–1.1[25]	6–7.5[23]
RBC	1.098[26]	2.63 (oblate spheroid)

# PBMC preparation – Real practice



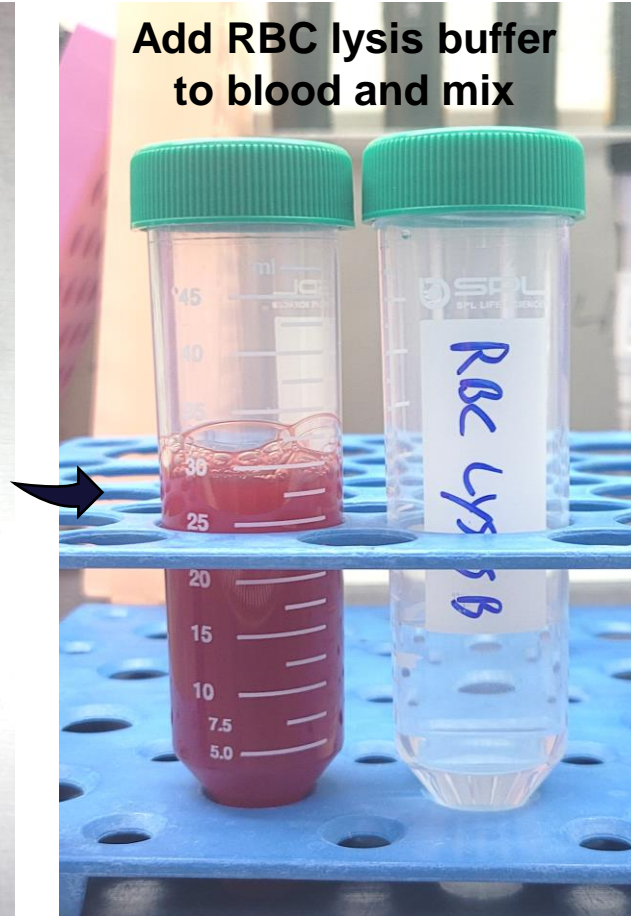
*Cell culture hood*

# Real practice – Whole Blood Lysis

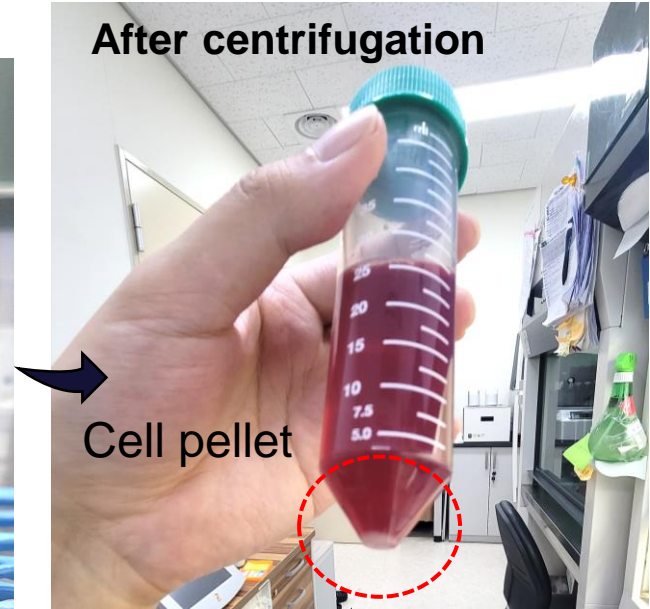
Patient blood



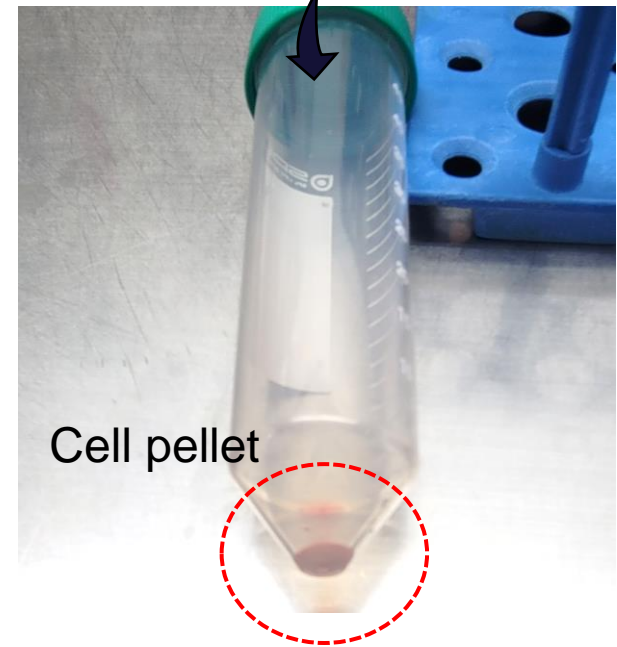
Add RBC lysis buffer to blood and mix



After centrifugation



Cell pellet

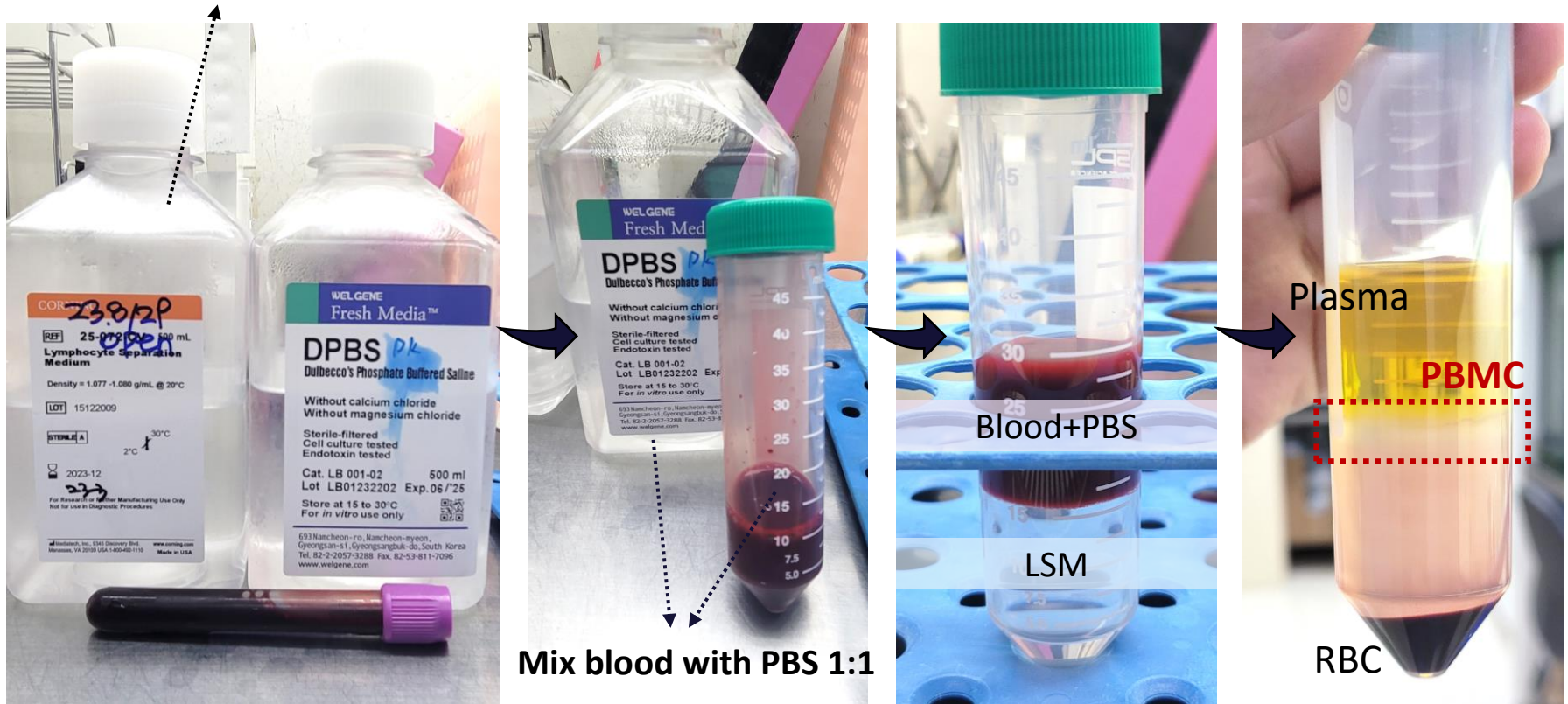


Cell pellet

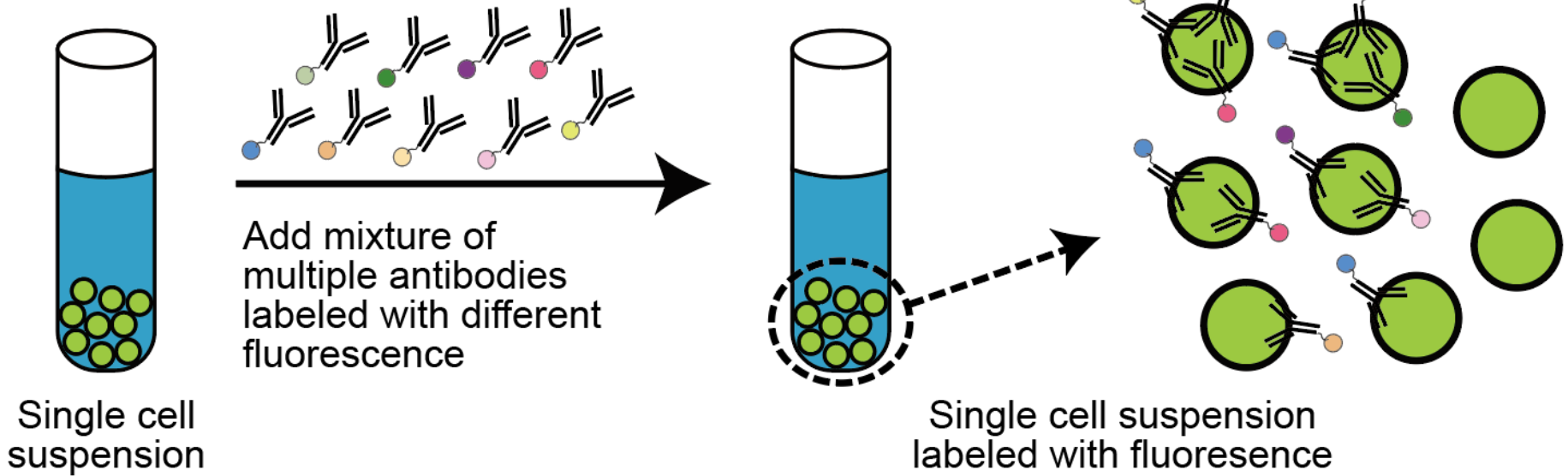
# Real practice – PBMC isolation

## Density gradient medium

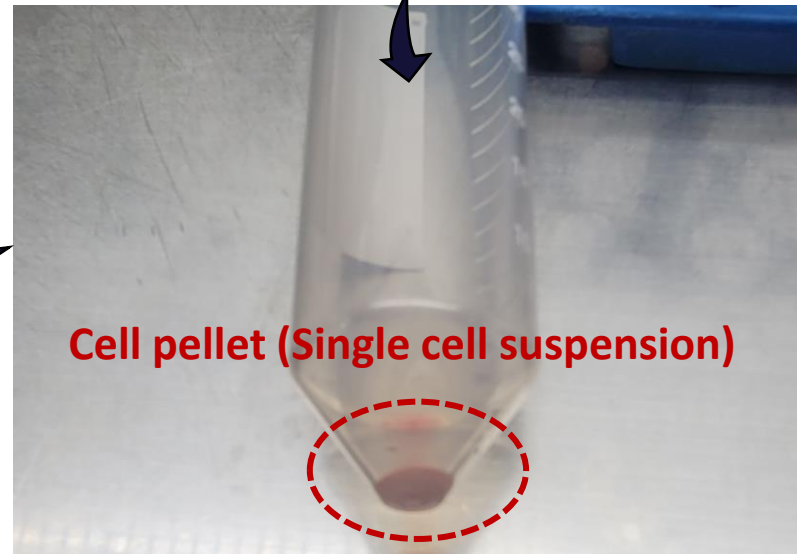
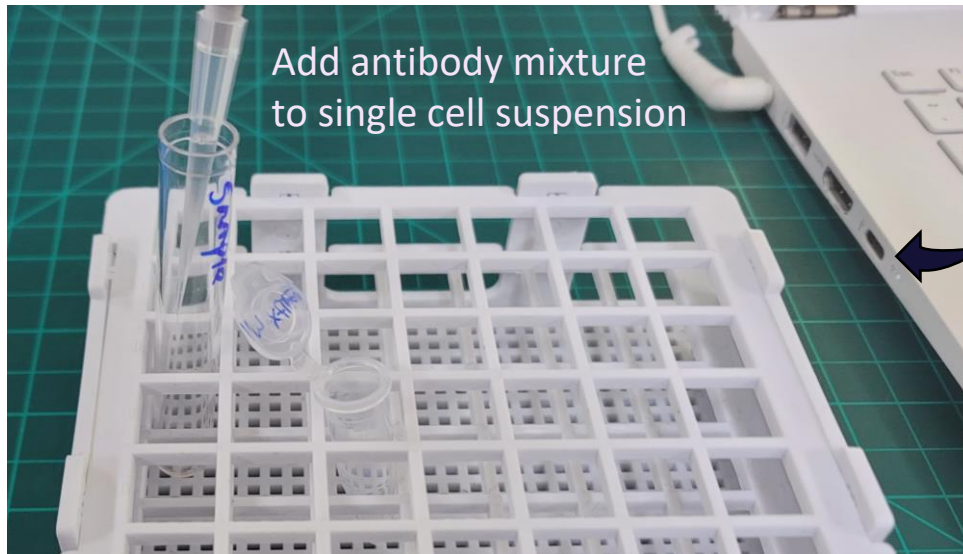
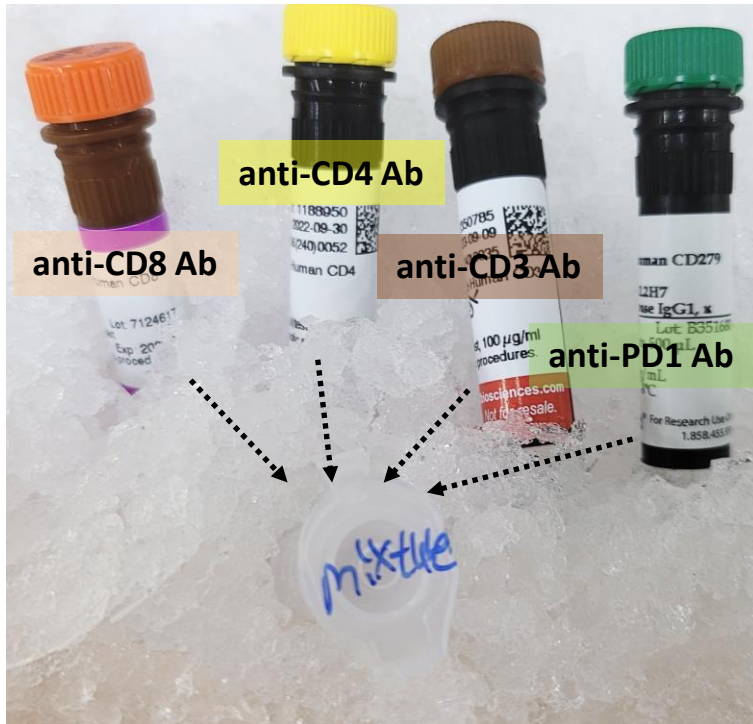
(Lymphocyte separation medium, LSM)



# FACS staining

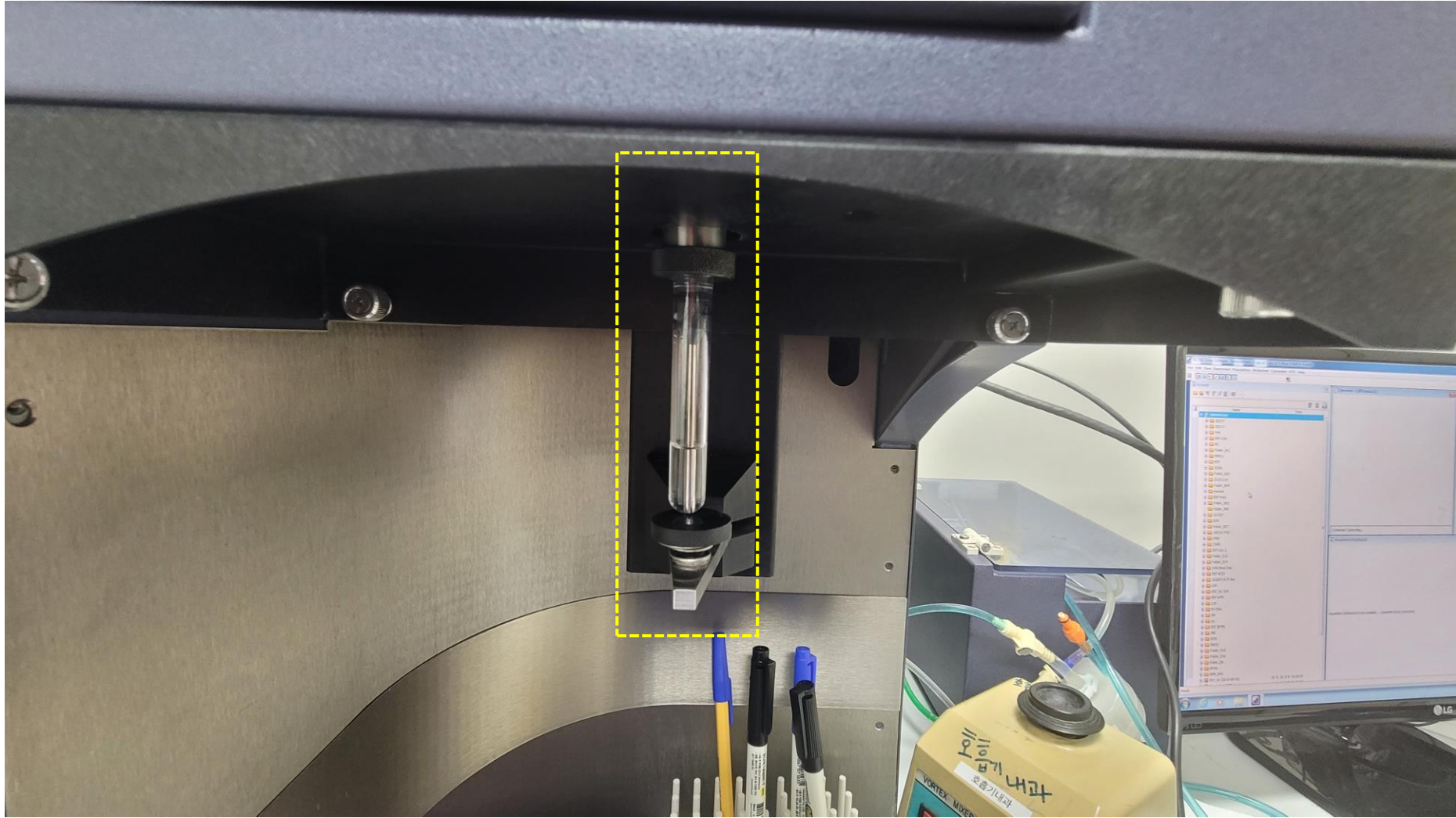


# FACS staining

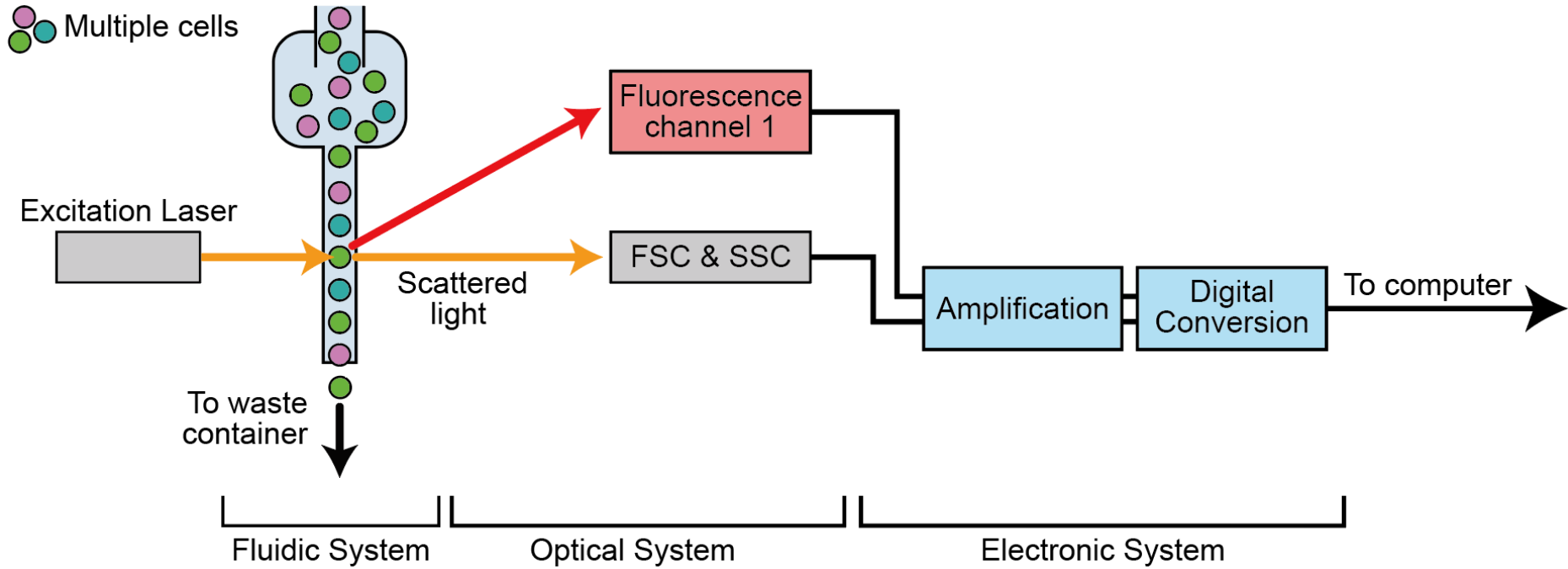




# 유세포 분석 기계



# 유세포 분석 기계



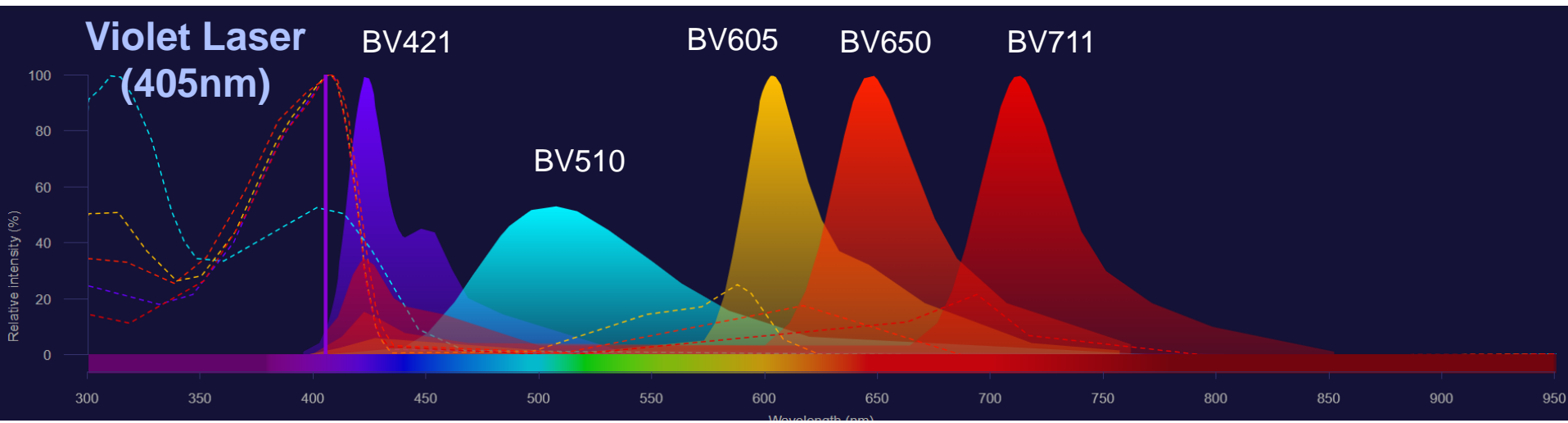
	Violet Laser (405nm)				
	BV421	BV510	BV605	BV650	BV711
Molecule			CD8	CD4	

	Blue laser (488nm)				
	FITC	PE	PE-TR	PerCP-Cy5.5	PE-Cy7
Molecule	PD-1				

	Red laser (640nm)		
	APC	A700	APC-Cy7
Molecule			CD3

# 유세포 분석 기계

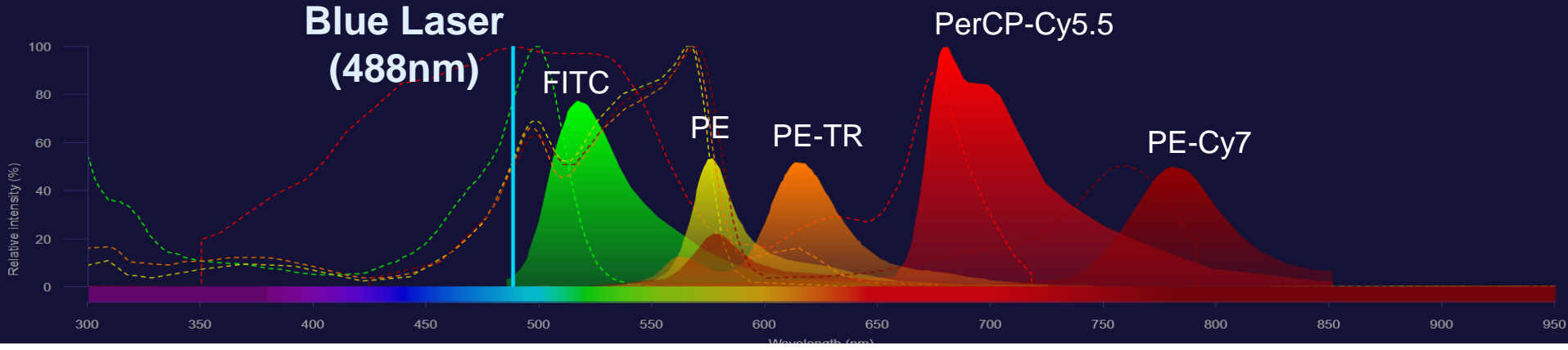
	Violet Laser (405nm)				
	BV421 (421nm)	BV510 (512nm)	BV605 (602nm)	BV650 (647nm)	BV711 (711nm)
Molecule			CD8	CD4	



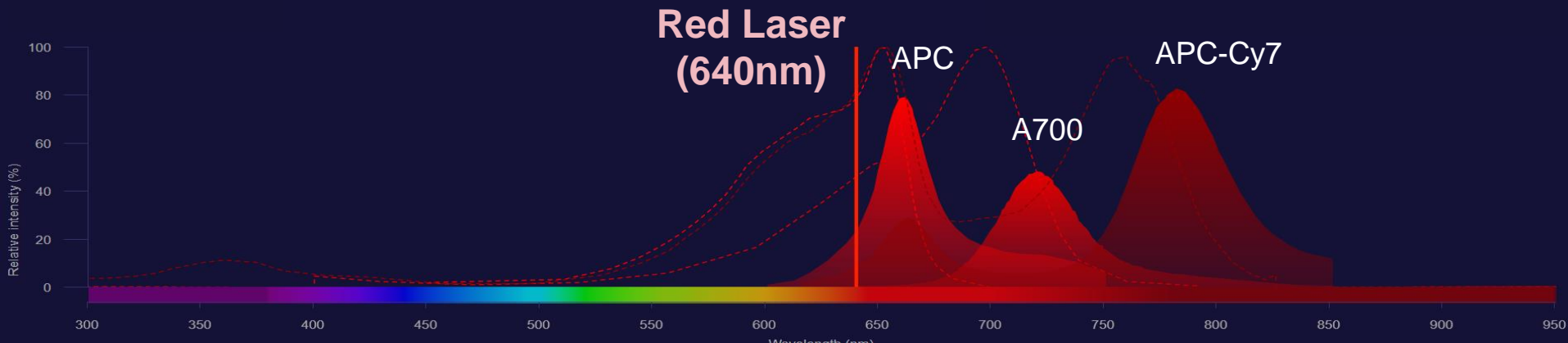
→ Violet Laser에 의해 excitation 되어 602nm에서 maximum emission을 보이는 형광이 결합된 **CD8**에 결합하는 항체와,  
 Violet Laser에 의해 excitation 되어 647nm에서 maximum emission을 보이는 형광이 결합된 **CD4**에 결합하는 항체를 사용해 유세포 분석을 하겠다.

# 유세포 분석 기계

	Blue laser (488nm)				
	FITC (516nm)	PE	PE-TR	PerCP-Cy5.5	PE-Cy7
Molecule	PD-1				



	Red laser (640nm)		
	APC	A700	APC-Cy7 (779nm)
Molecule			CD3



# 유세포 분석 기계

The screenshot displays the FACSDiva software interface. On the left is a file browser showing a tree structure of folders and experiments. The main window is divided into several panels:

- Parameters Panel:** Shows a table of parameters for FSC and SSC.
- Acquisition Dashboard:** Displays current activity for Tube\_001, including Threshold Rate (0 evt/s), Stopping Gate Events (0 evt), and Elapsed Time (00:00:00). It also includes basic controls like Next Tube, Acquire Data, Record Data, and Restart.
- Worksheet:** A large grid area on the right labeled "Worksheet - 실제 기록된 유세포 데이터가 보이는 곳".

Overlaid text boxes provide additional context:

- Cytometer - 어떤 형광을 확인할지** (Cytometer - Which fluorescence to check)
- Dashboard - 유세포 기계 조정** (Dashboard - Flow cytometer adjustment)

Parameter	Voltage	Log	A	H	W
FSC	333	<input type="checkbox"/>	<input checked="" type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
SSC	241	<input type="checkbox"/>	<input checked="" type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>

Active Tube/Well	Threshold Rate	Stopping Gate Events	Elapsed Time
Tube_001	0 evt/s	0 evt	00:00:00

# 유세포 분석 기계

• BV650	480	<input checked="" type="checkbox"/>	<input checked="" type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
• FITC	450	<input checked="" type="checkbox"/>	<input checked="" type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
• APC-Cy7	580	<input checked="" type="checkbox"/>	<input checked="" type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>

Add      Delete

Cytometer Connected

## Acquisition Dashboard

Current Activity

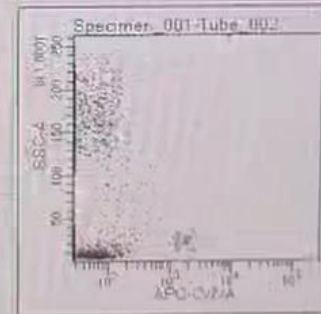
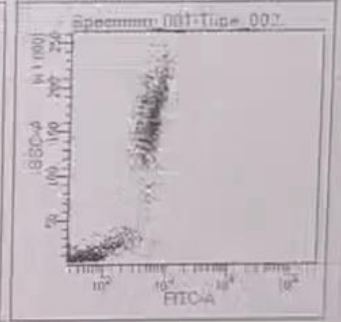
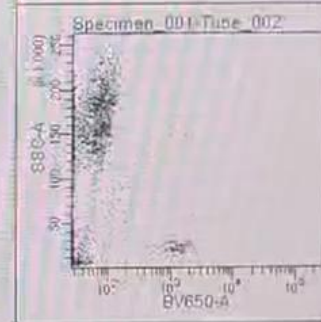
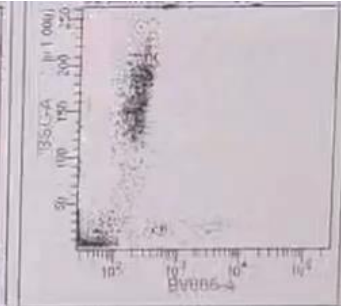
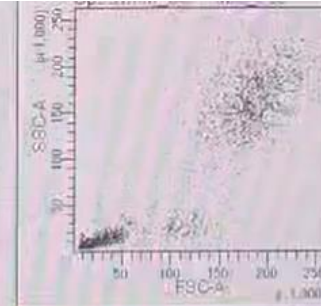
Active Tube/Well	Threshold Rate	Stopping Gate Events	Elapsed Time
Tube 002	3324 evt/s	0 evt	00:00:01

Basic Controls

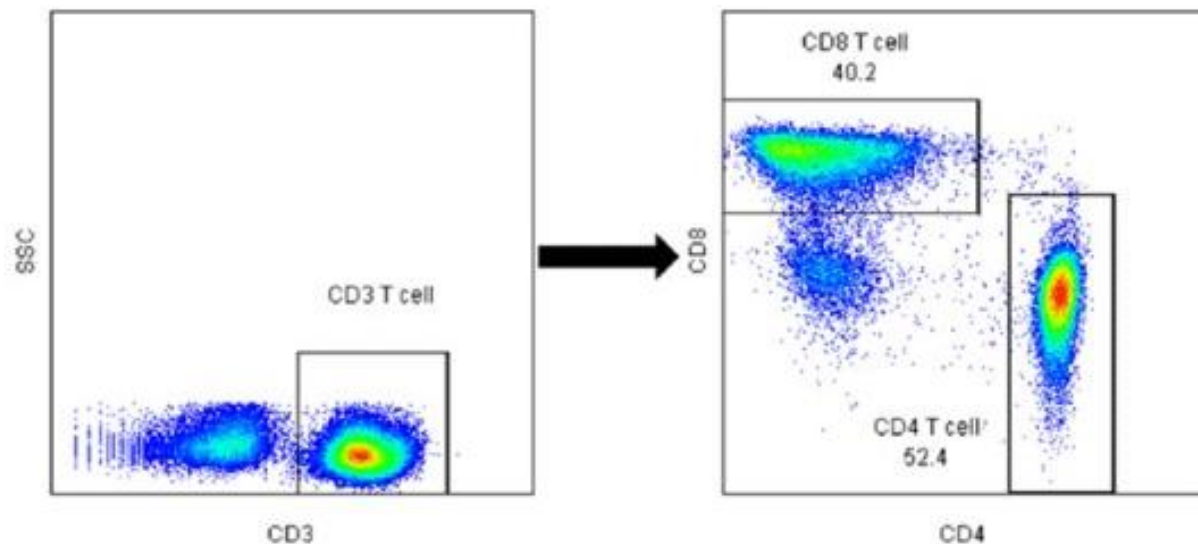
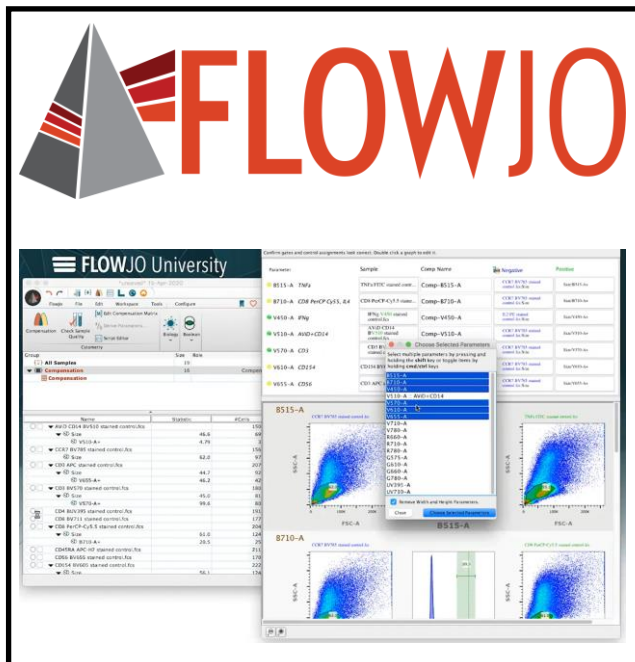
Acquisition Setup

Stopping Gate:  All Events    Events To Record: 100000 evt    Stopping Time (sec): 0

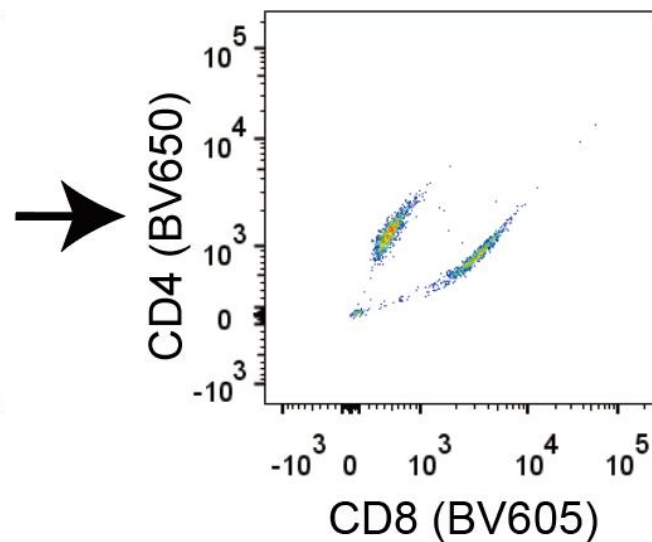
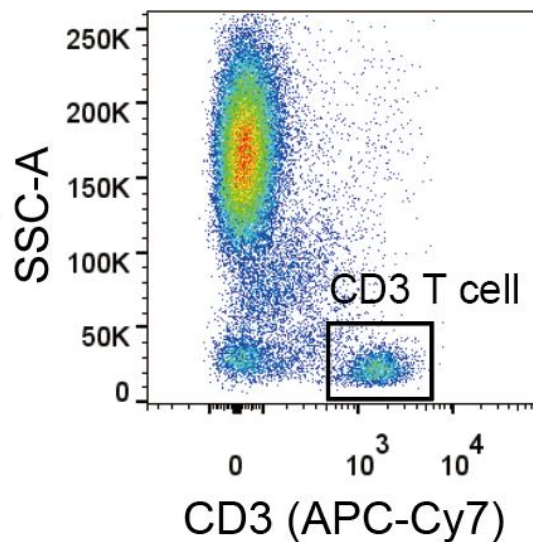
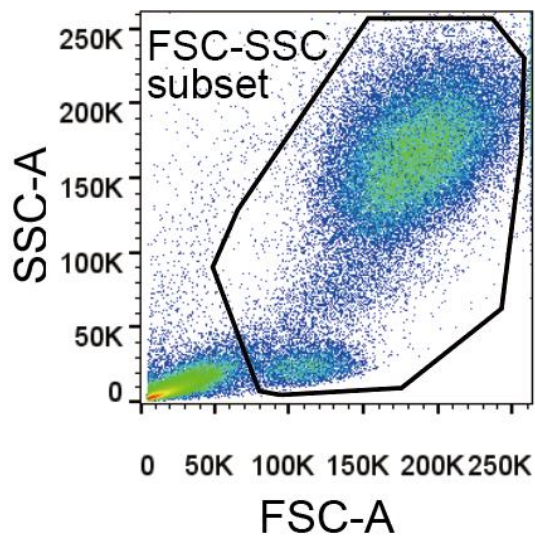
Storage Gate:  All Events    Events To Display: 100000 evt



# 유세포 분석 기계

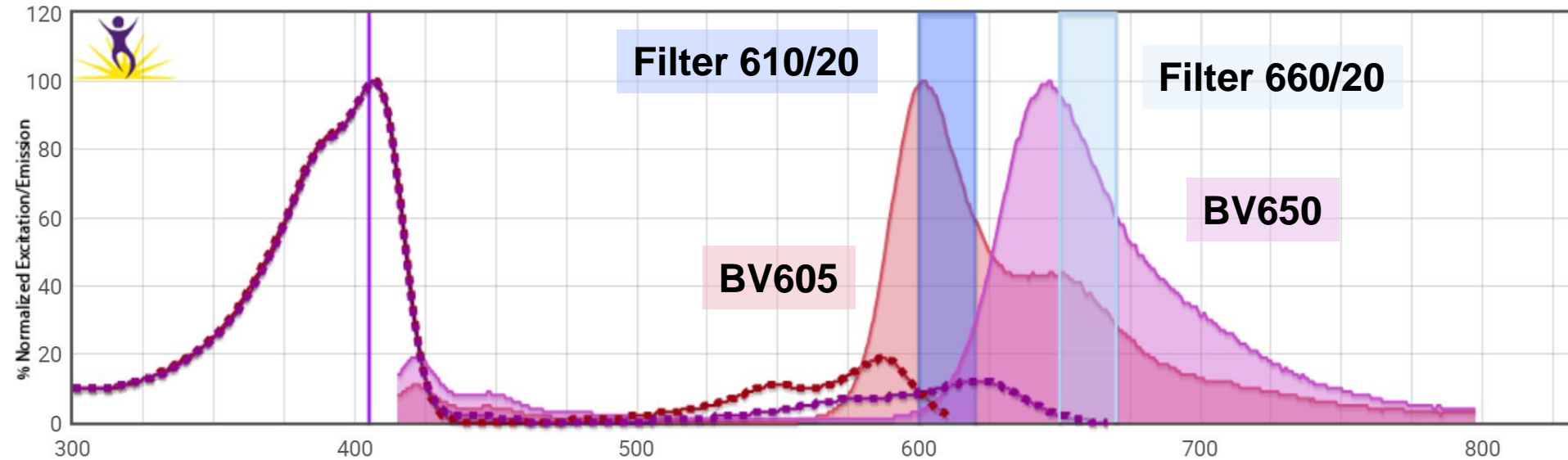


*Xiaoyan et al., 2019, BMC Nephrology*



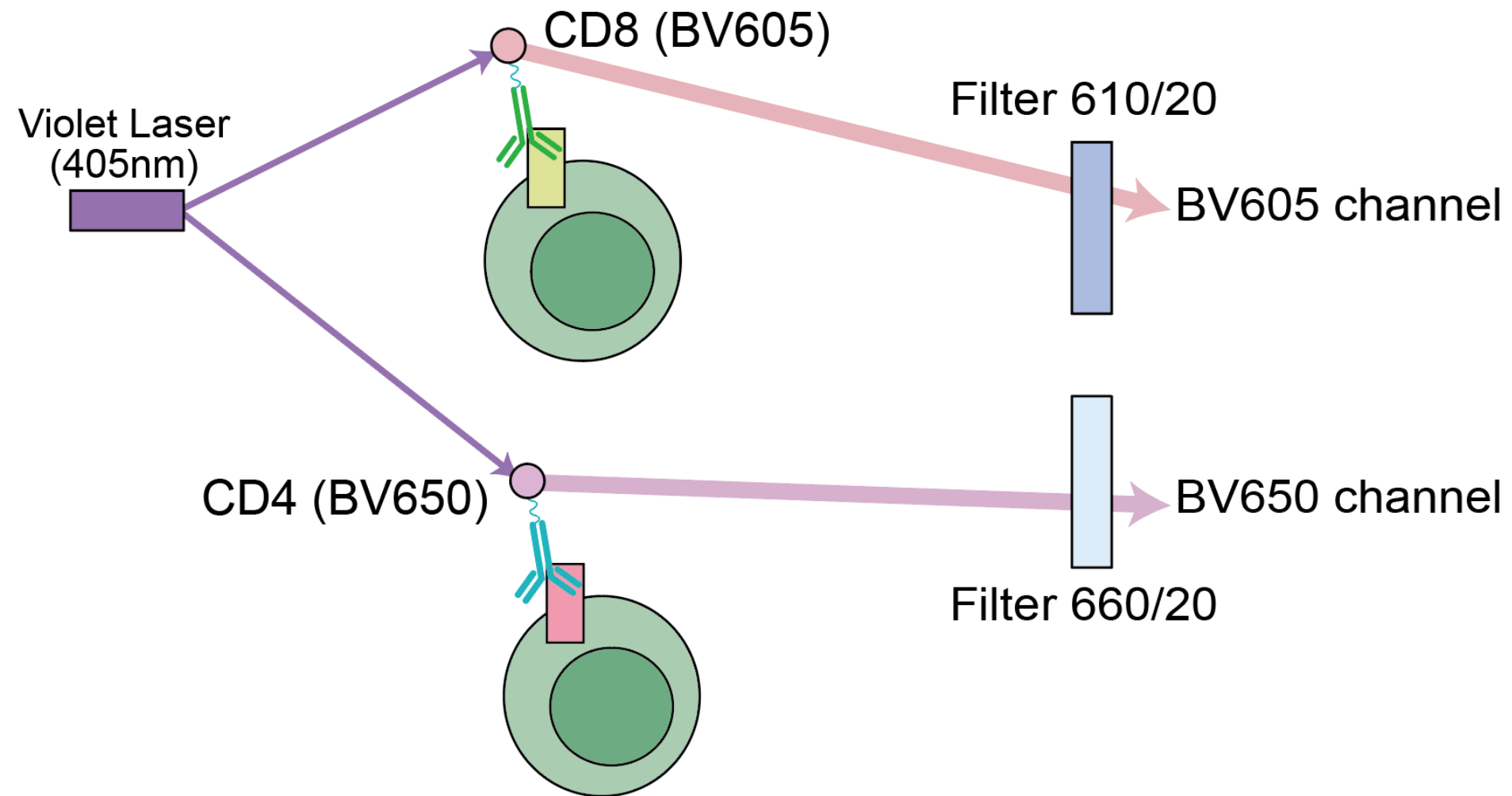
# Compensation

	Violet Laser (405nm)				
	BV421	BV510	BV605	BV650	BV711
Molecule			CD8	CD4	

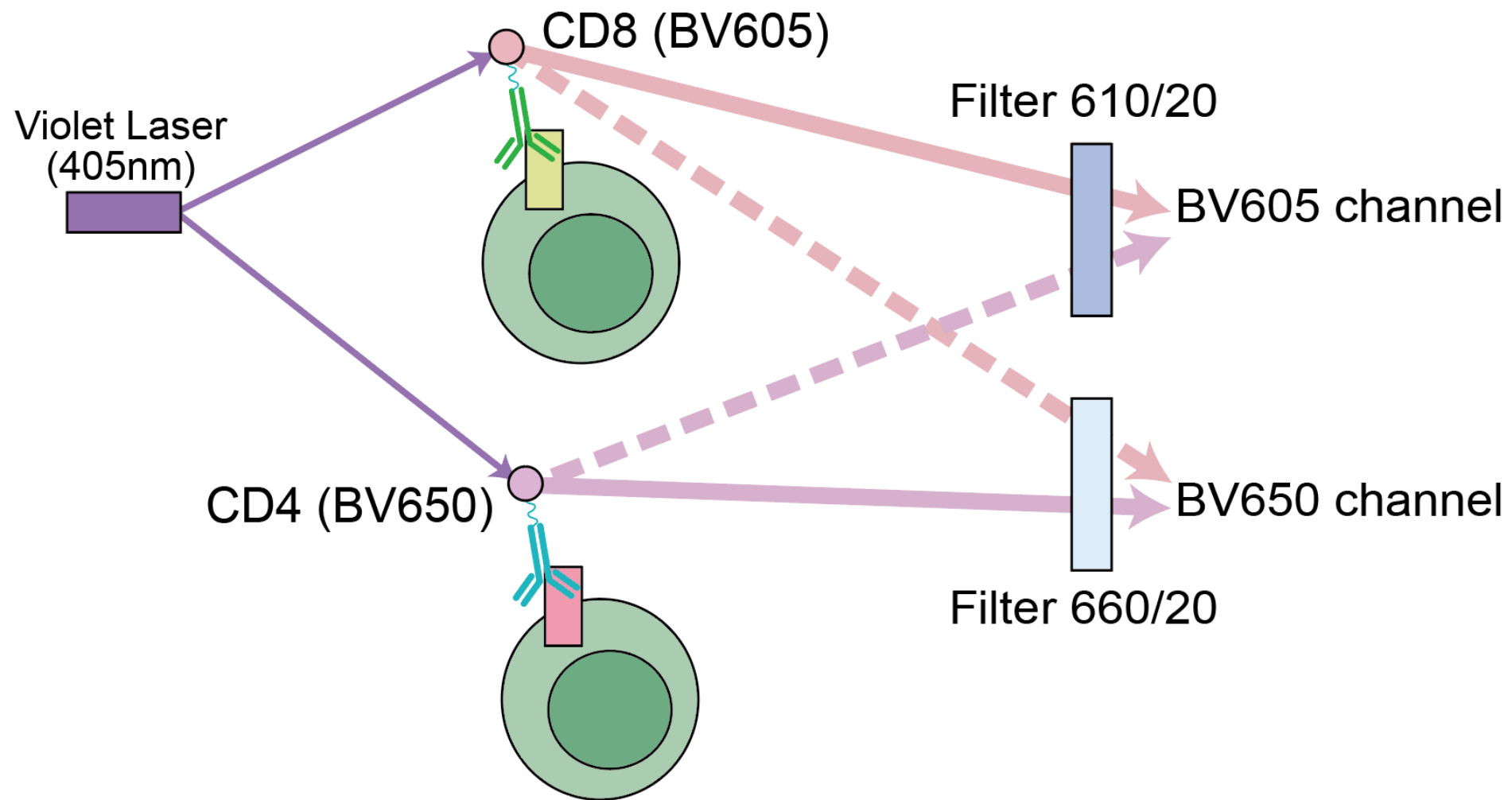


각 파장이 서로 침범하여 겹치는 현상을 'Spill over'라고 한다.  
Spill over를 보정해주는 과정을 Compensation 이라 한다.

# Compensation



# Compensation



# Compensation

Cytometer - LSRFortessa (1)

Status Parameters Threshold Laser Compensation Ratio

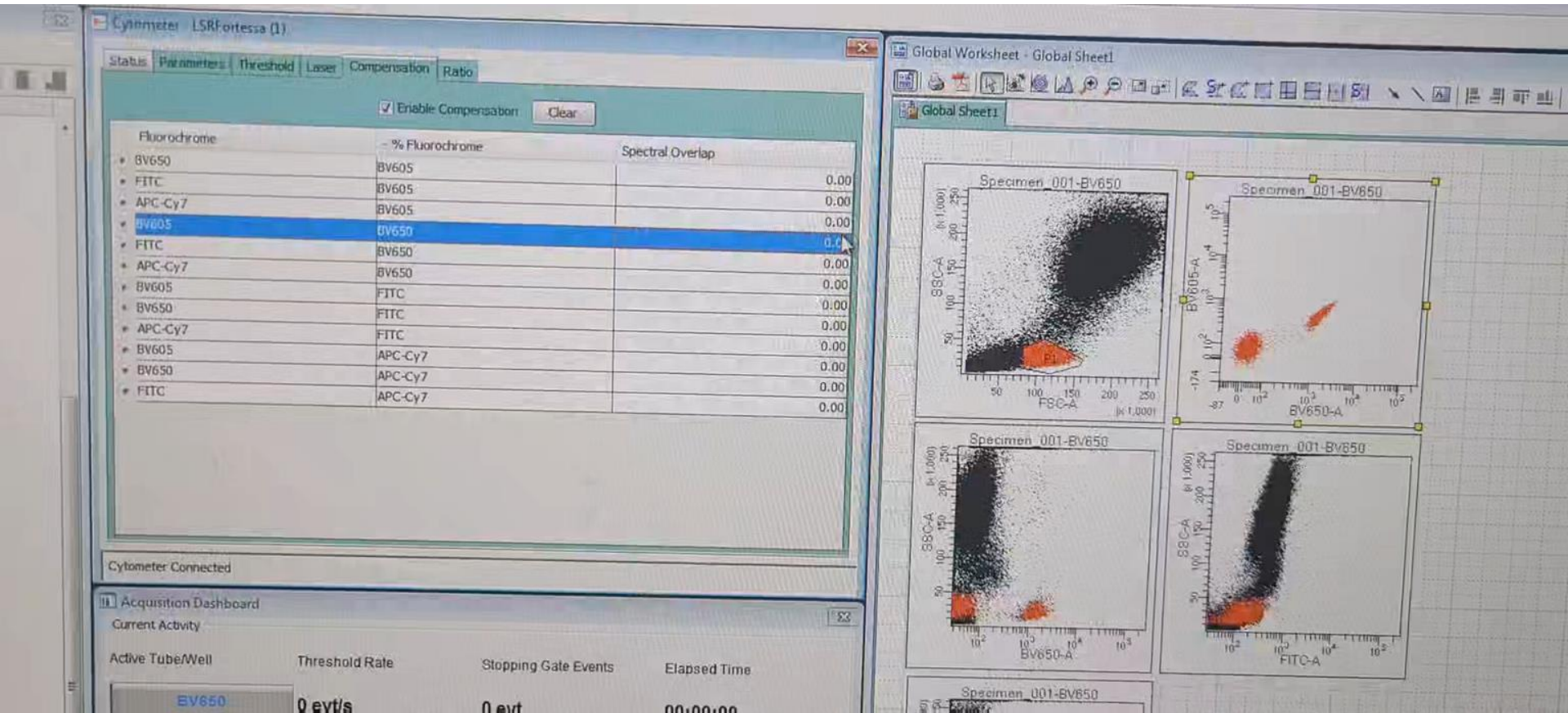
Enable Compensation

Clear

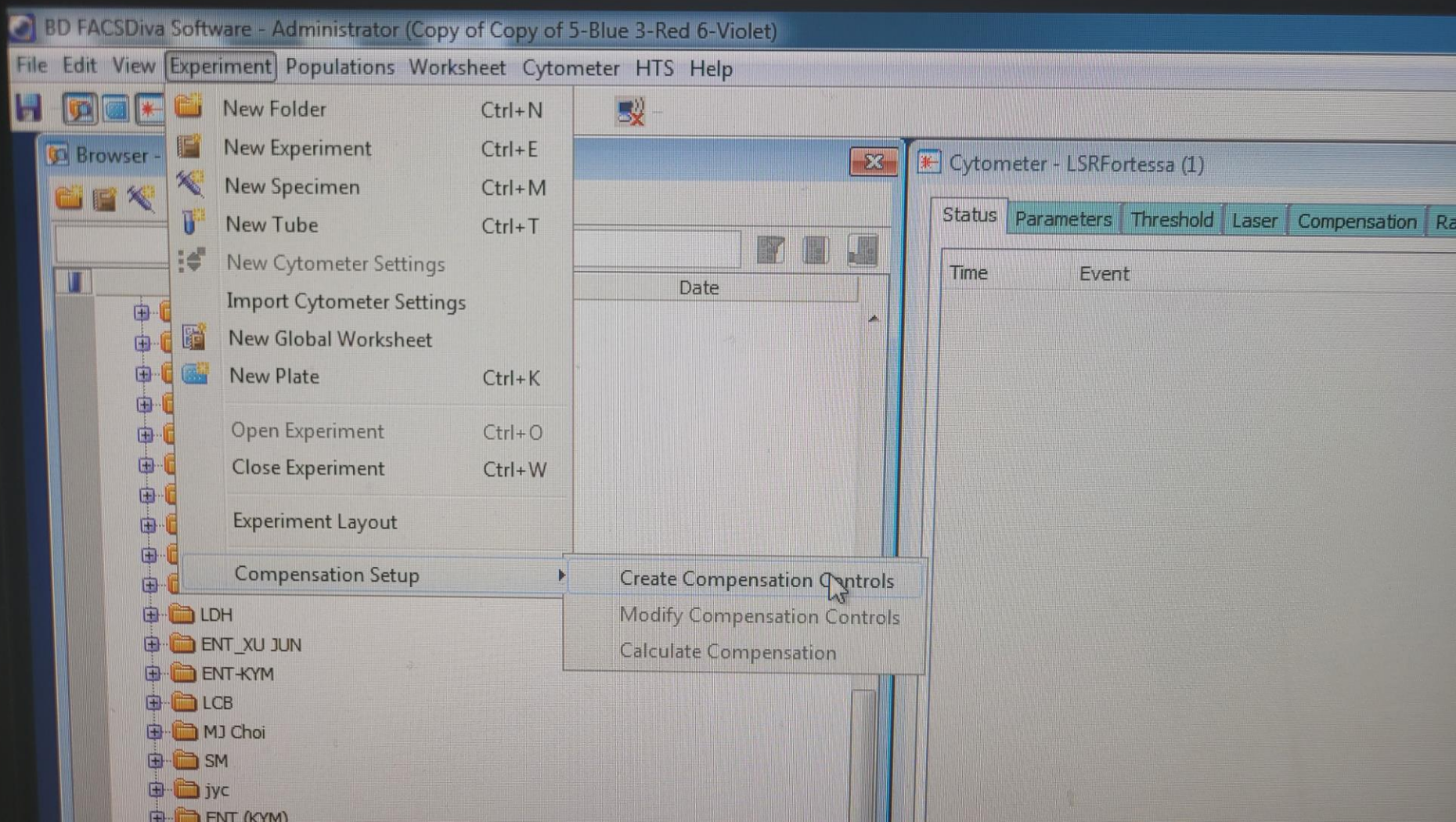
Fluorochrome	- % Fluorochrome	Spectral Overlap
BV650	BV605	0.00
FITC	BV605	0.00
APC-Cy7	BV605	0.00
BV605	BV650	0.00
FITC	BV650	0.00
APC-Cy7	BV650	0.00
BV605	FITC	0.00
BV650	FITC	0.00
APC-Cy7	FITC	0.00
BV605	APC-Cy7	0.00
BV650	APC-Cy7	0.00
FITC	APC-Cy7	0.00

**BV605 channel 에서 BV650 color 와의 spectral overlap 만큼을 빼준다.**

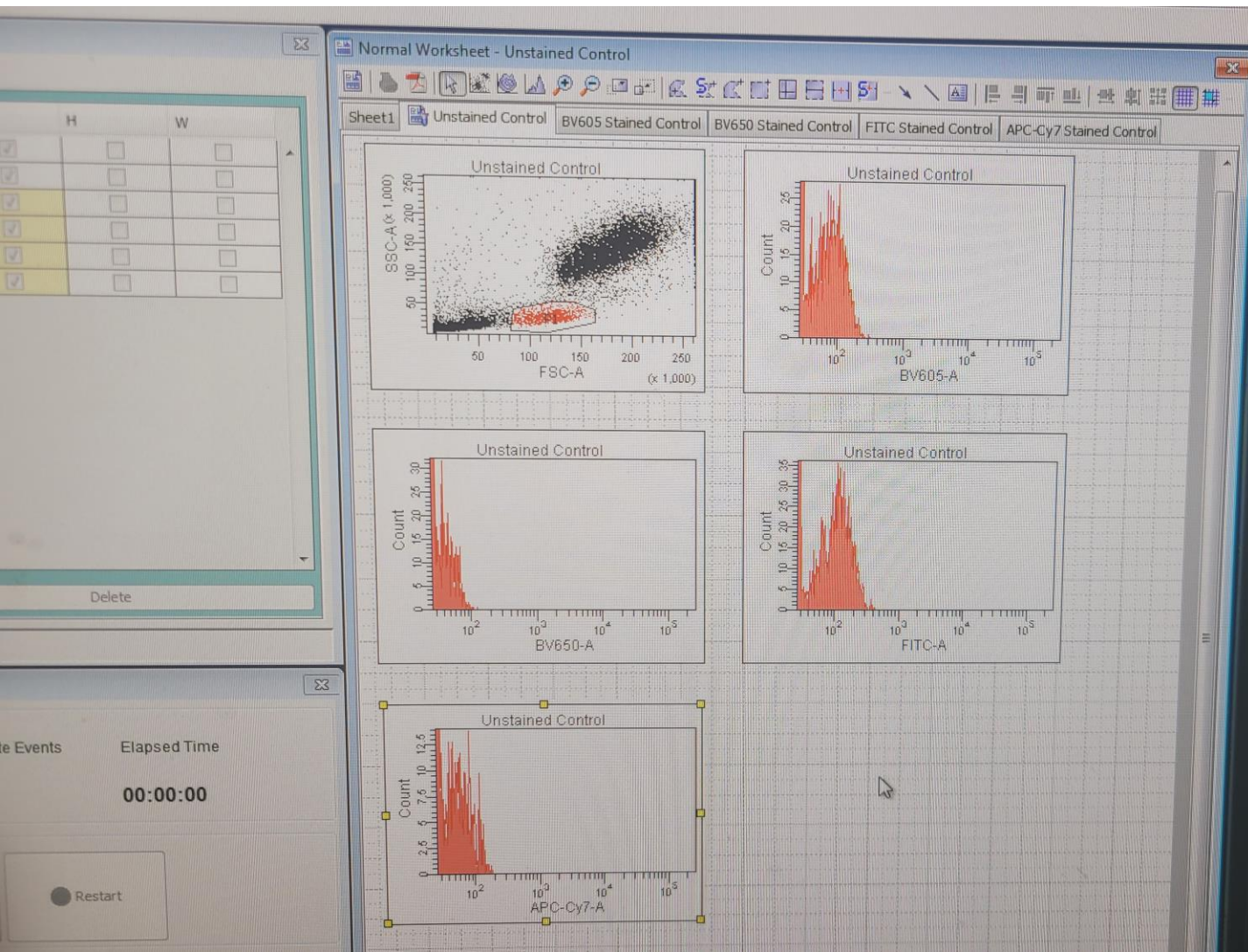
# Compensation



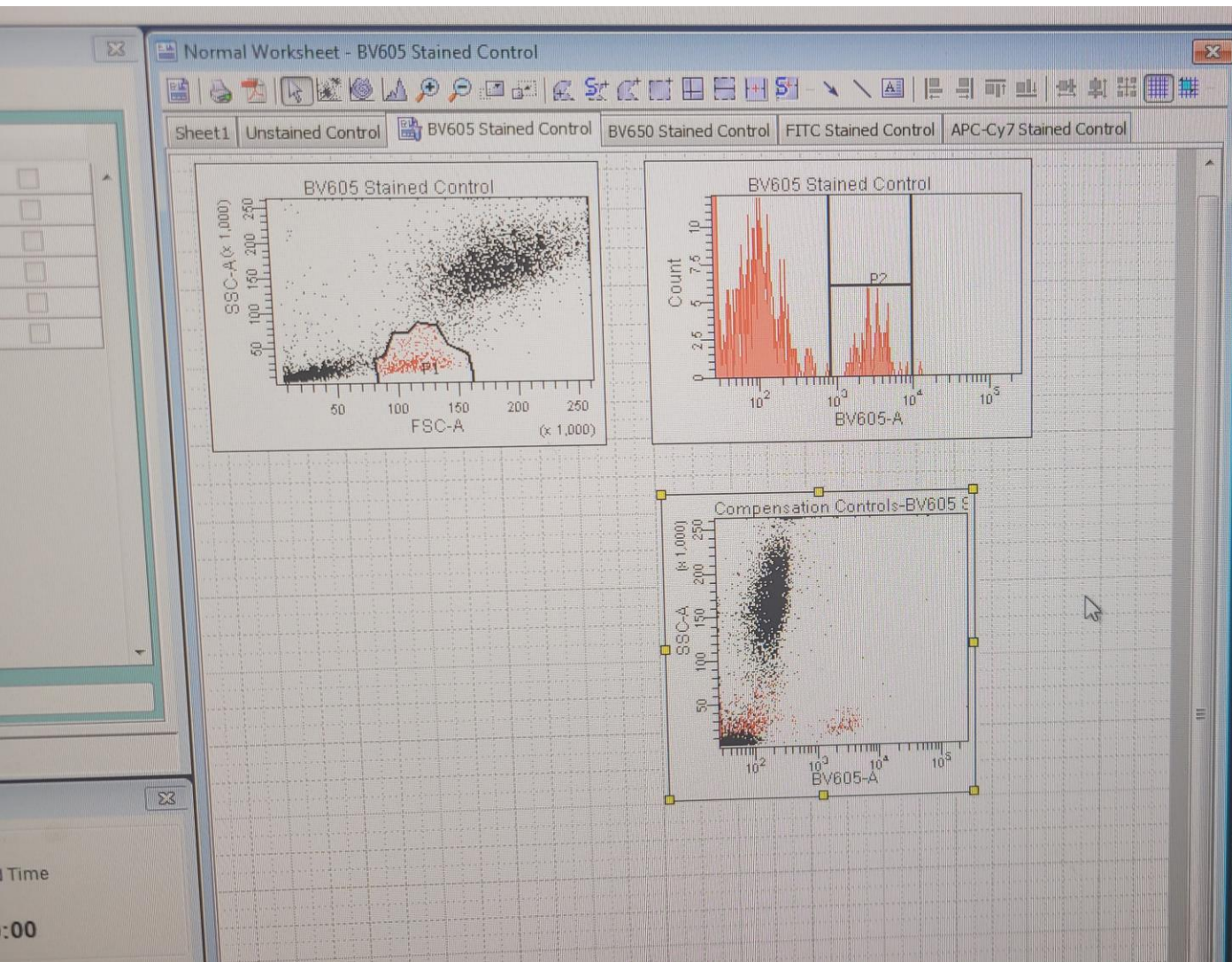
# Compensation



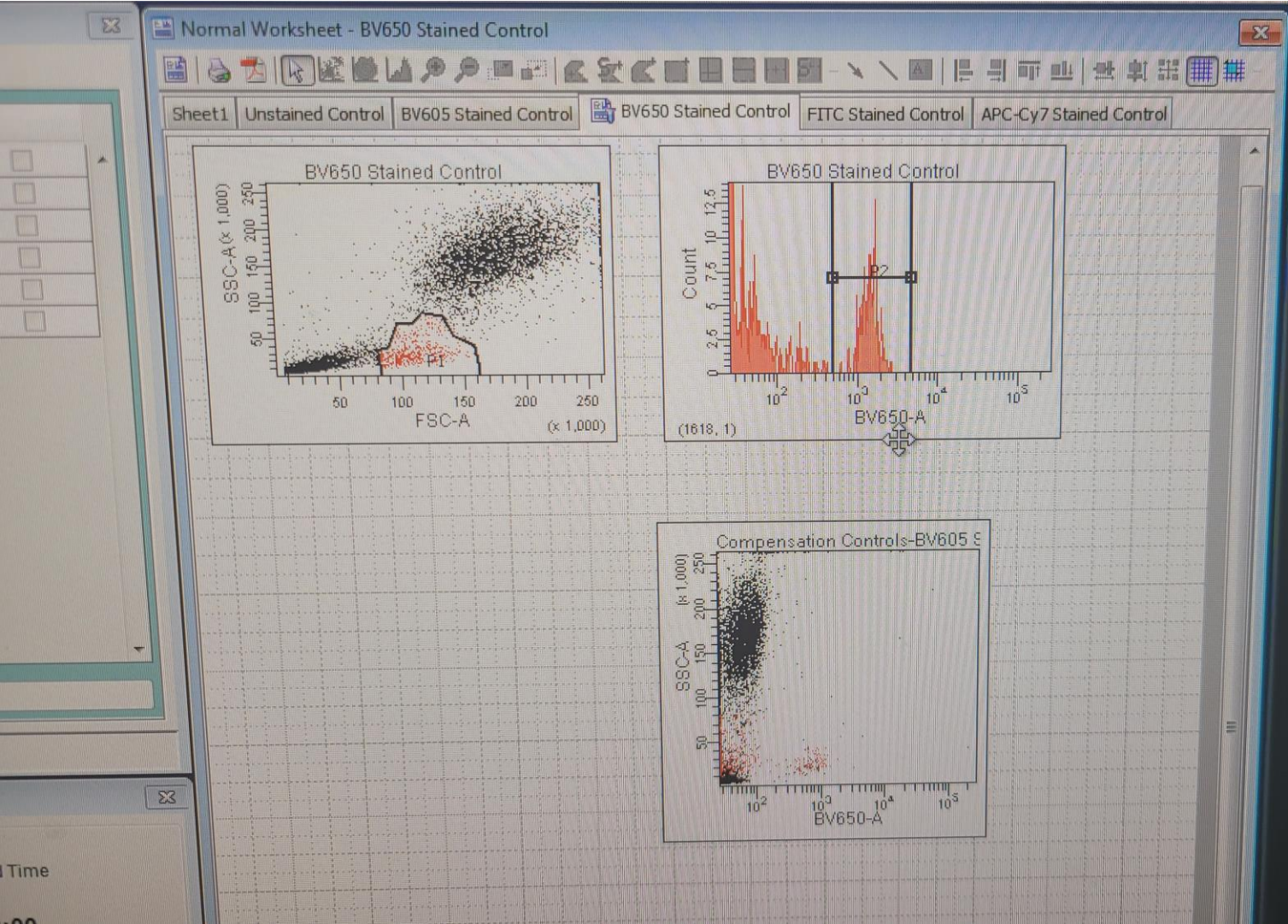
# Compensation



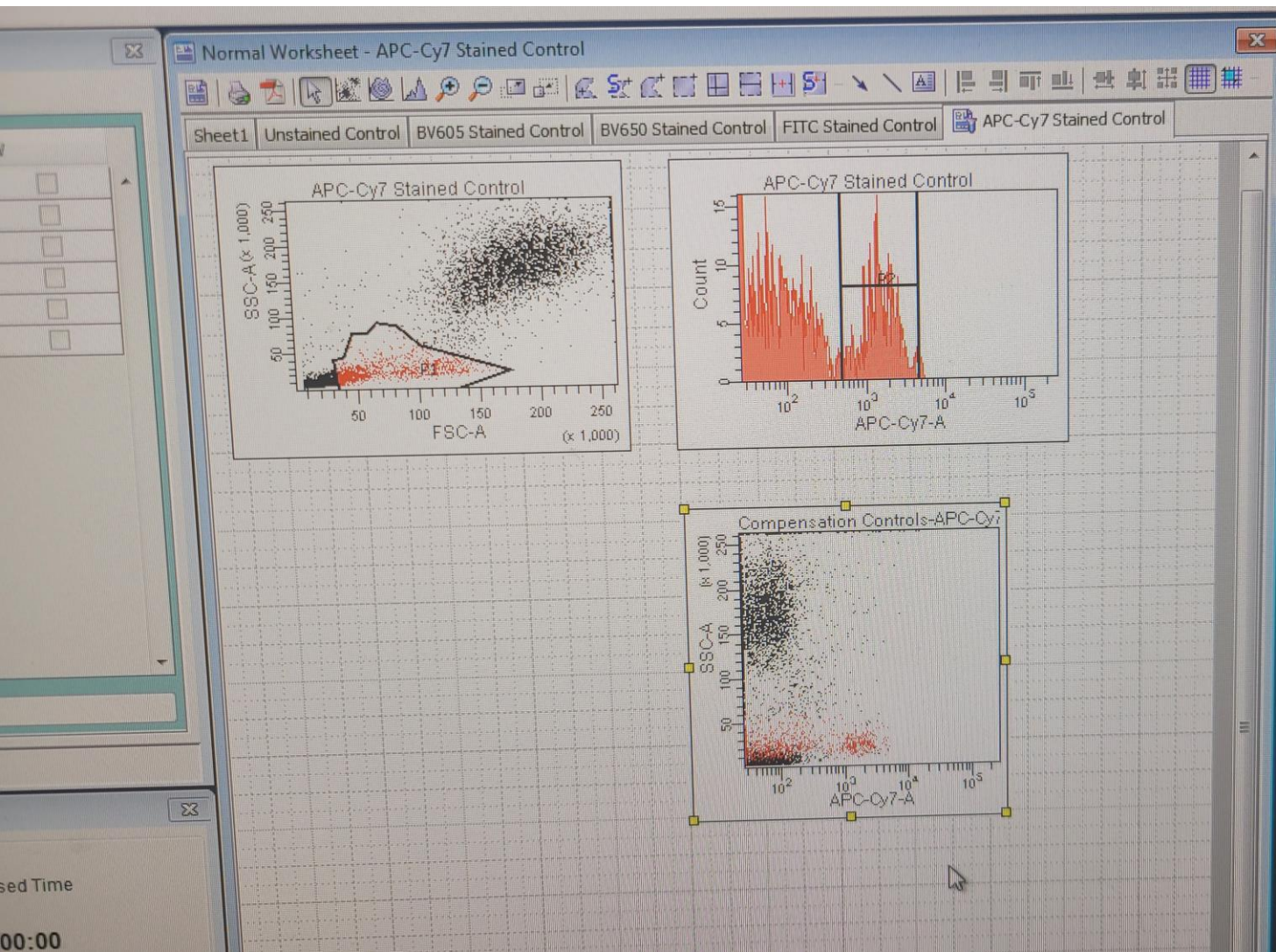
# Compensation



# Compensation



# Compensation

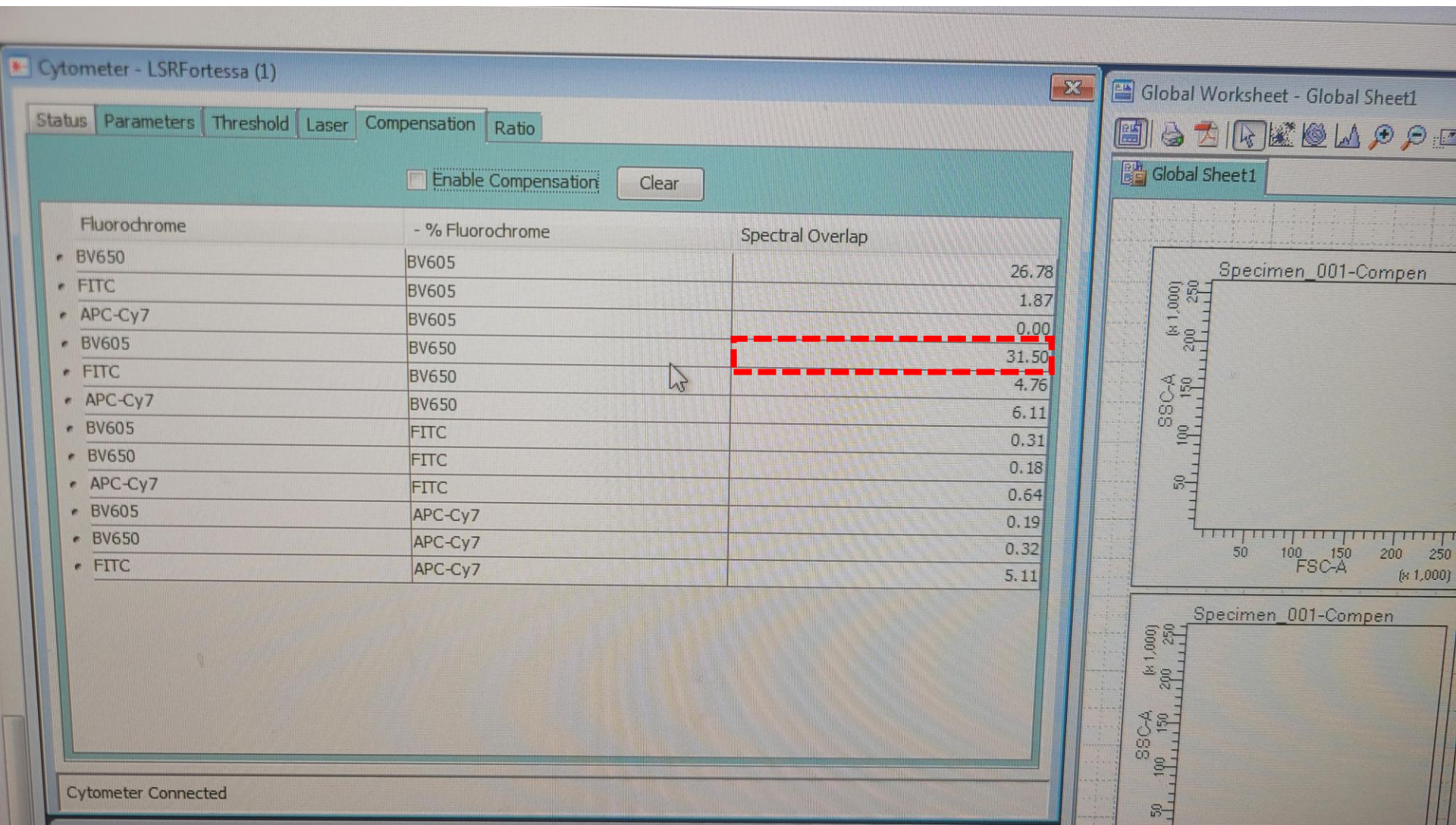


# Compensation

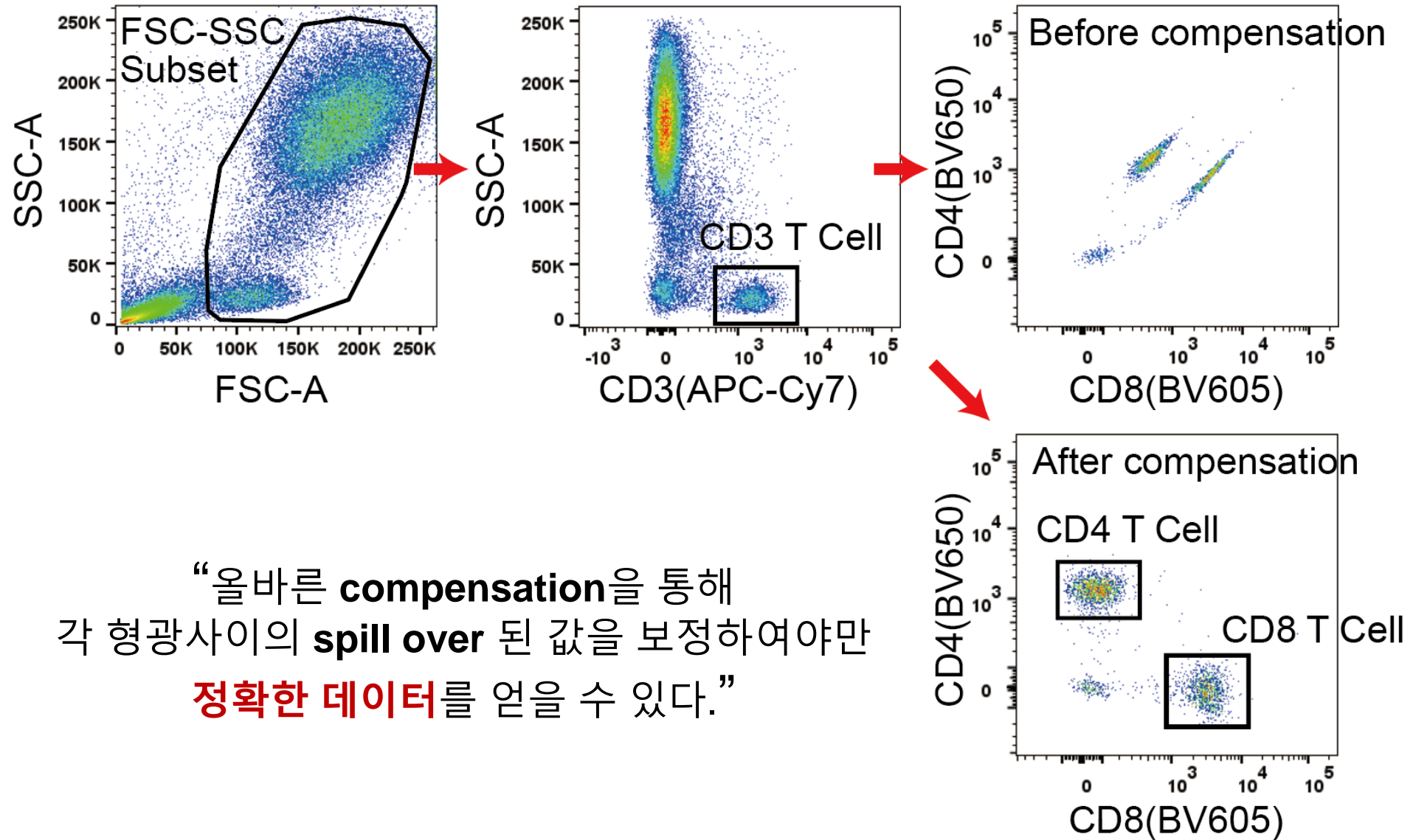
The screenshot displays the BD FACSDiva Software interface. On the left, the 'Experiment' menu is open, showing options like 'New Folder', 'New Experiment', and 'Compensation Setup'. The 'Compensation Setup' sub-menu is expanded, highlighting 'Calculate Compensation'. On the right, the 'Cytometer - LSRFortessa (1)' window is open, showing the 'Compensation' tab with a table of parameters.

Parameter	Voltage	Log	A
FSC	150	<input type="checkbox"/>	<input checked="" type="checkbox"/>
SSC	235	<input type="checkbox"/>	<input checked="" type="checkbox"/>
BV605	480	<input checked="" type="checkbox"/>	<input checked="" type="checkbox"/>
BV650	450	<input checked="" type="checkbox"/>	<input checked="" type="checkbox"/>
FITC	450	<input checked="" type="checkbox"/>	<input checked="" type="checkbox"/>
APC-Cy7	580	<input checked="" type="checkbox"/>	<input checked="" type="checkbox"/>

# Compensation



# Compensation



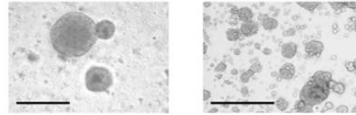
“올바른 **compensation**을 통해  
각 형광사이의 **spill over** 된 값을 보정하여야만  
**정확한 데이터**를 얻을 수 있다.”

# 담배 추출물을 활용한 실험예시



control, 0.25%, 0.5%, 1% 흡연 추출물 노출

폐 오가노이드 변화 분석



No. of organoids  
Organoid growth rate  
Size of organoids

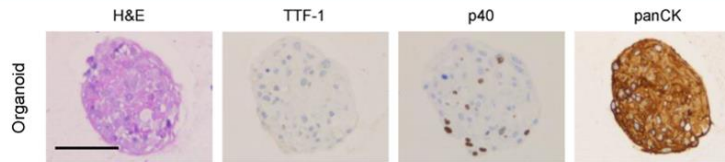
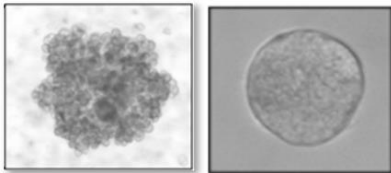


충남대학교병원  
정재욱 교수님

흡연과 관련된 폐암  
오가노이드 모델 구축

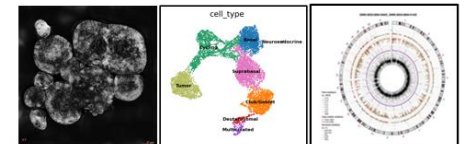
폐암 특이적 바이오  
마커 발굴

폐 오가노이드



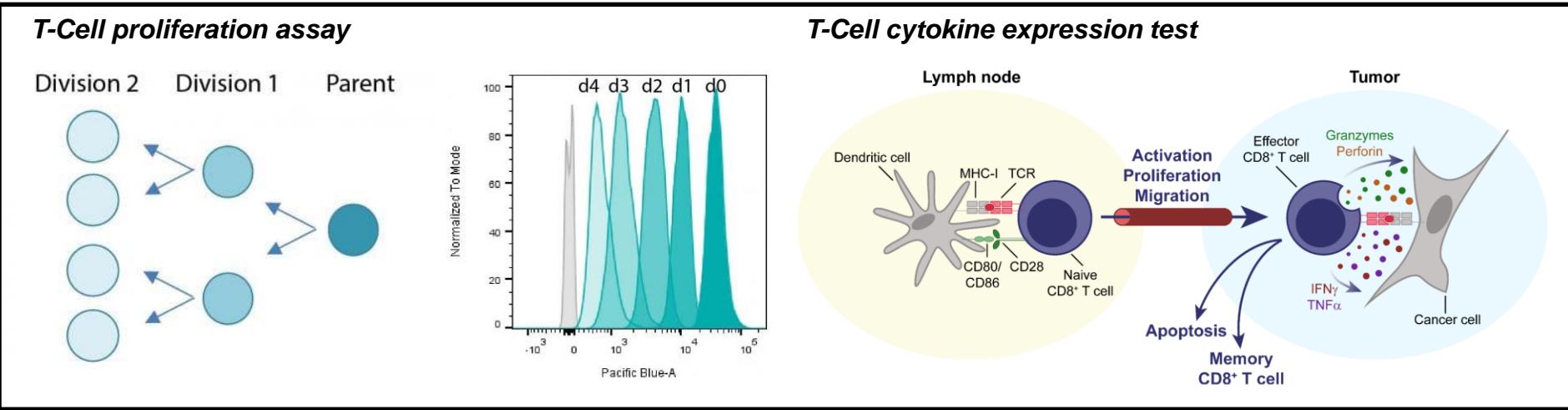
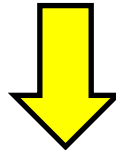
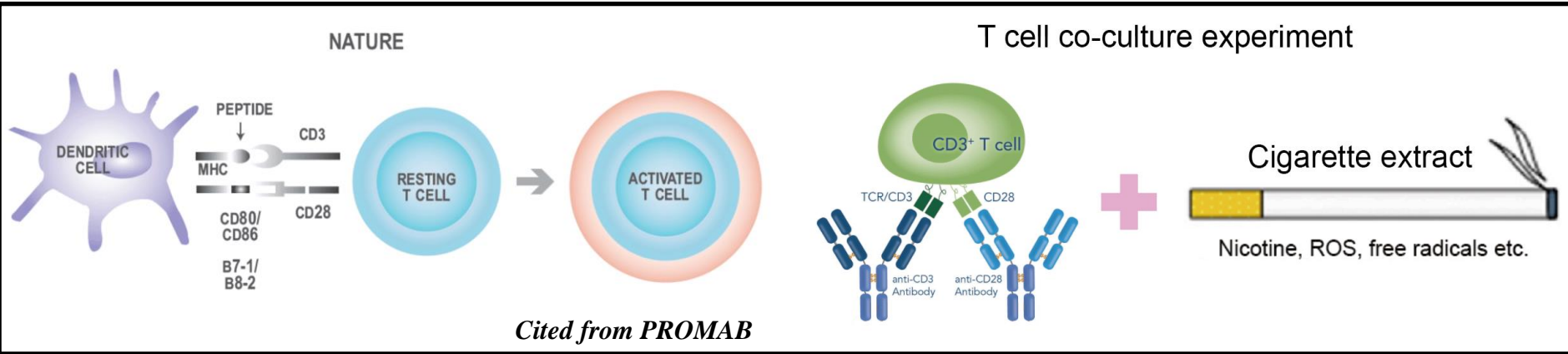
폐/폐암 오가노이드의 구조 및 단백질/유전자 발현 분석

- H & E, Western Blot, 면역염색, Microarray 분석,  
3D 이미징, 단세포 RNA 분석, NGS 검사를 바탕으로  
흡연 추출에 노출된 폐 오가노이드의 유전자 및 단백질 발현 확인



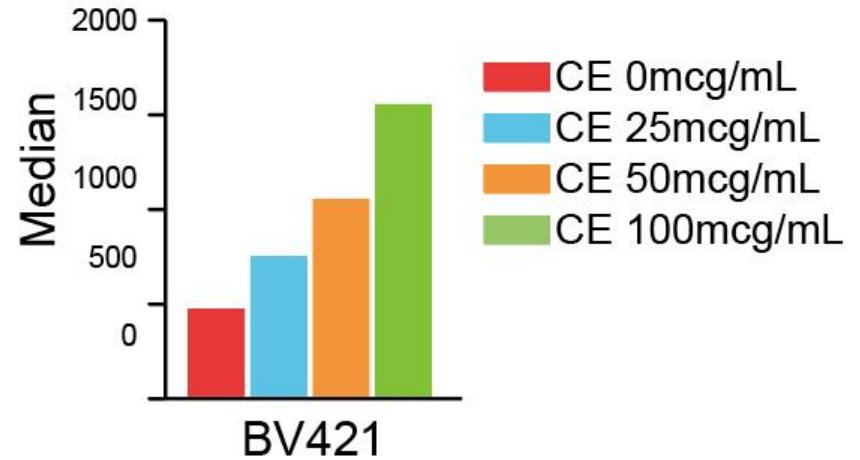
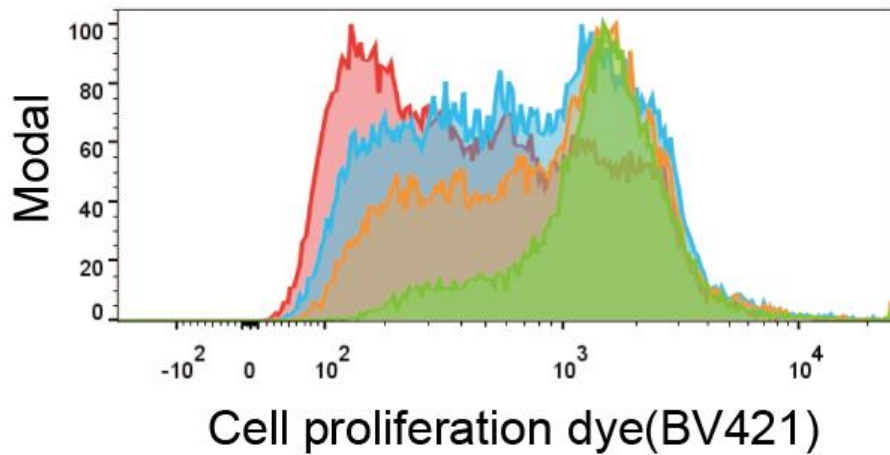
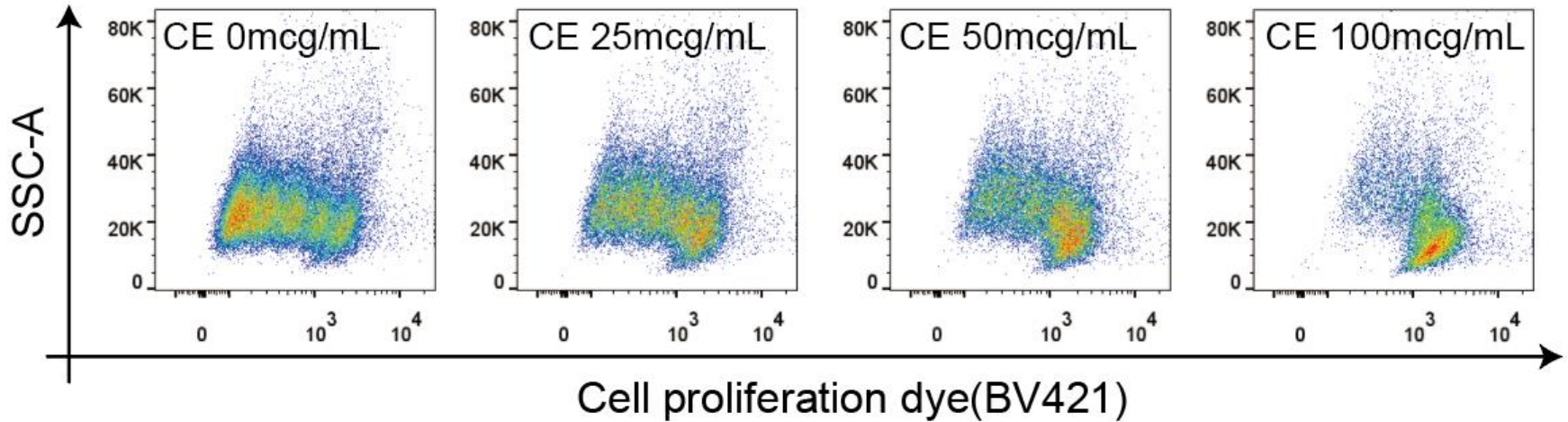
공동 배양 기술 개발

# 담배 추출물을 활용한 실험예시



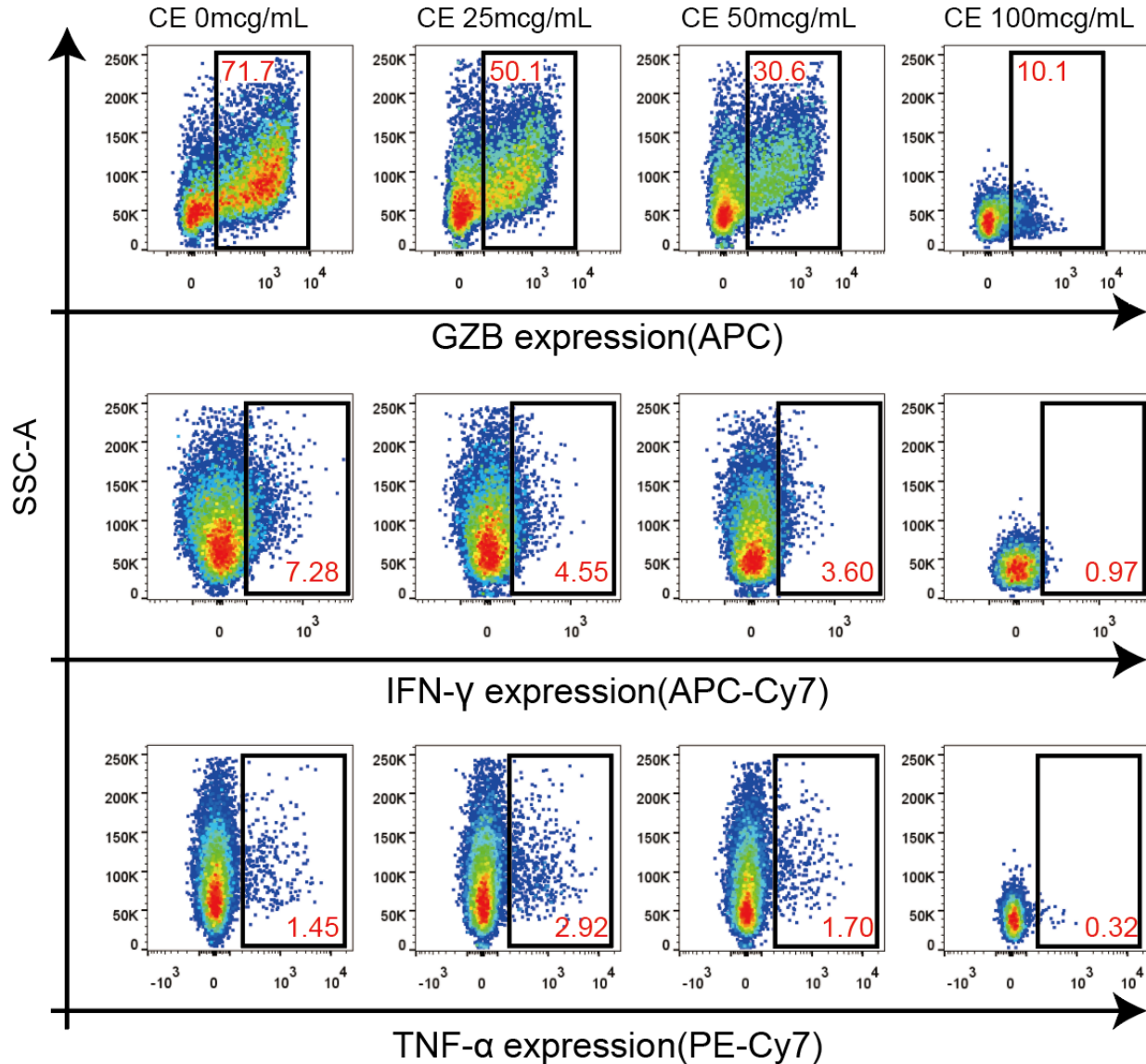
# 담배 추출물을 활용한 실험예시 – T cell proliferation assay

\*\*CE : Cigarette extract



# 담배 추출물을 활용한 실험예시 - T cell cytokine expression test

\*\*CE : Cigarette extract



# Acknowledgement



*Prof. Dong-il Park*



*Prof. Chaek Chung*

***Thank you  
for listening***



***Department Respiratory and Critical Medicine in CNUH***